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3 **Leaf Chlorophylls and Carotenoids Status and**

4 **their correlation with storage root weight of**

5 **Some Local and Exotic Sweetpotato Genotypes**

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10 **ABSTRACT**

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The investigation was carried out to characterize of chlorophyll components and carotenoids of leaves of some local and exotic genotypes of sweetpotato namely Local-1, Local-2, Local-5, Local-8, Exotic-1, Exotic-2, Exotic-4 and BARI SP-4 and their effect on production of total dry matter and dry weight of storage roots during November 2015 to April 2016. The experiment was laid out in Randomized Complete Block Design with three applications. Fresh leaves of 5-6th position from the top of vine were collected from the research field into polybag with proper tagging to the laboratory at morning at 30, 60, 90 and 120 days after planting. Collected leaves were washed, wiped out of excess water, cut into small pieces leaving away mid ribs, mixed thoroughly, and 250 mg of leaf materials were taken in a mortar. Leaf materials were grinded finely by a pestle with 25 ml of cold 80% acetone for two minutes. Sample tubes were centrifuged for 10 minutes. The homogenate was filtered and made up to 25 ml with cold 80% acetone. The centrifuged samples were incubated in dark for half an hour. The optical density (OD) for each solution was measured at 663, 645 and 440.5 nm against 80% acetone as blank in one cm cell of spectrophotometer. Triplicate estimation was done for each character. Chemical analyses were performed at Regional Laboratory of Soil Resource Development Institute, Sylhet-3100, Bangladesh. Statistical analyses was done using MSTATC software following analysis of variance technique and Duncan's Multiple Range Test. Results show that chlorophyll-a gradually increased up to 60 DAP in all genotypes, thereafter it continued only in Exotic-4, Exotic-3 and Local-1 up to 90 DAP. The highest amount of chlorophyll-a ( $10.27 \pm 0.45$  mg 100 gfw<sup>-1</sup>) was in Local-1 at 90 DAP. The highest amount of chlorophyll-b (mg 100 gfw<sup>-1</sup>) was in Exotic-3 ( $19.13 \pm 0.53$ ) followed by Local-1 ( $16.85 \pm 0.50$ ) at 30 DAP. Carotenoids content of all genotypes increased gradually up to 90 DAP and thereafter decreased except Exotic-4. The highest carotenoids was in Exotic-3 ( $10.78$  mg 100 gfw<sup>-1</sup>) followed by Local-1 ( $10.13$  mg 100 gfw<sup>-1</sup>) at 90 DAP. At 120 DAP, the highest storage roots weight was in Local-8 ( $232.40 \pm 5.97$ ), followed by Local-1 ( $187.50 \pm 5.23$ ). Chlorophylls and carotenoids had no significant effect on total dry matter and storage roots dry weights at 30 DAP. All chlorophyll components and carotenoids had positive correlation with TDM and storage roots dry weights at 120 DAP. In conclusion, chlorophyll-a had positive effect on storage roots dry weight after 90 DAP while chlorophyll-b came to positive effect on storage roots dry weights after 60 DAP. At final harvest, the higher chlorophyll, carotenoids and dry matter synthesizing genotypes were Local-1 and Local-8.

12 **Keywords:** *Chlorophyll, carotenoids, exotic genotype, local genotype, yield, correlation*

13

14 **1. INTRODUCTION**

15

16 Sweetpotato (*Ipomoea batatas* L. Lam.) is a dicotyledonous plant of the family

17 Convolvulaceae. It is perennial in nature but it is grown in an annual crop. The plants bear

18 adventitious roots which enlarge near the stem and form edible storage roots. In Bangladesh

19 it is produced about 0.761 million ton on about 0.045 million ha of land with an average yield  
20 of 16.91 t ha<sup>-1</sup> [1]. It is characterized by low production, yield and storage root quality  
21 compared to Japan, Senegal and Israel [2]. However, it is easy to grow and capable of  
22 growing under adverse weather and soil conditions. It requires low input and less  
23 management practices [3]. It is a very efficient food crop and produces more dry matter,  
24 protein and minerals per unit area in comparison to cereals [4]. Storage roots are rich source  
25 of energy, several minerals and micronutrients [5] and leaves are rich in vitamin B, beta-  
26 carotene, iron, calcium, zinc and protein [6].

27  
28 The yield of sweetpotato depends on the production of assimilates (source) and its  
29 accumulations (sink). Storage roots (number and weight) are predominant sink whereas  
30 leaves and tender vines are the source. The photosynthetic rate and the leaf area are  
31 regarded as the source potential. Leaves take part in the production of assimilates. The  
32 leaves of plant contain chlorophylls (Chlorophyll-a, Chlorophyll-b) and carotenoids.  
33 Chlorophyll-a possesses a green-blue color while chlorophyll-b possesses green-yellow  
34 color [7]. Chlorophyll with the pigments has a central role in light harvesting, photosystem  
35 protection, and other growth functions [8, 9]. Carotenoids participate in harvesting light  
36 energy for photosynthesis [10]. They are also involved in the defense mechanism against  
37 oxidative stress [11], and play an essential role in the dissipation of excess light energy and  
38 provide protection to reaction centers [12, 13].

39  
40 Recent studies show that chlorophyll and carotenoids have positive effect on human health.  
41 Chlorophyll is often referred to as the green blood of plants due to the identical molecular  
42 structure with hemoglobin with only difference in center atom (iron or magnesium). This  
43 similarity makes chlorophyll so important to our health, it improve digestive, immune and  
44 detoxification systems of human body [14]. Leaves contain phenols, flavonoids,  $\beta$ -carotene,  
45 anthocyanin, and caffeoylquinic acid derivatives [15]. Carotenoids extract from leaves  
46 functions as a cheap natural yellow dye. It can be beneficial to human health compare to the  
47 artificial colouring dye [16].

48  
49 There are many local sweetpotato genotypes are available in Sylhet region and many of  
50 them are growing at the farmer's level sporadically. Rajput *et al.* [17] reported that the  
51 functional leaves may directly reflect to yield. Besides, for adaptive trial of improved  
52 sweetpotato cultivars developed by different countries and organization, it is necessary to  
53 determine chlorophyll and carotenoids. Moreover, it is necessary to determine how these  
54 genotypes/cultivars can be made available to the farmers of Bangladesh. Therefore, an  
55 experiment was undertaken to characterize different types of chlorophyll features of leaves  
56 of local and exotic genotypes of sweetpotato.

## 57 **2. METHODOLOGY**

58 The experiment was carried out at Sylhet Agricultural University Farm, Tilagarh, Sylhet  
59 during November 2015 to March 2016. It lies between 24°54'33.5" to 24°54'34.7" N latitude  
60 and 91°54' 04.6" to 91°54'05.6" E longitude. Texture of top soils was Loam (sand 50%, silt  
61 37%, clay 13%). Well drained soil. Top soil (0-15 cm) is deep brown in color. The soil is  
62 characterized as Ramgarh soil series of Northern an Eastern Piedmont Plains (AEZ 22) in  
63 Bangladesh [18].

### 64 **2.1 Planting Materials**

65 Nine sweetpotato genotypes/cultivars namely Local-1, Local-2, Local-5, Local-8, Exotic-1,  
66 Exotic-2, Exotic-4 and BARI SP-4 were used as planting materials while BARI SP-4 was  
67 check variety.

### 68 **2.2 Experimental Procedure and Design**

69 The experiment was set in a Randomized Complete Block Design (RCBD) with three  
 70 replications. The experimental field was fertilized with manures and fertilizers as per soil test  
 71 value: Cowdung =5000 kg, Urea =212 kg, TSP =186 kg, MoP =187 kg, Gypsum =63 kg,  
 72 Zinc sulfate (Hepta) =9 kg, Solubor =3 kg, Magnesium sulfate =84 kg and dolomite = 988 kg.  
 73 Before final land preparation, half of urea and MoP, full of other fertilizers and cow dung  
 74 were applied. Rest of Urea and MoP were applied as side dressing after 35 days of planting  
 75 at earthing up operation. Soil reaction (pH) was corrected by dolomite application prior to 15  
 76 days of planting followed by bed preparation. Raised beds were prepared and cuttings of  
 77 sweetpotato vines were planted in lines maintaining row to row 60 cm and plant to plant 30  
 78 cm. The unit plot size was of 4.8 m x 4.2 m with a block to block distance 1.0 m and plot to  
 79 plot distance 0.6 m. Weeding was done as and when necessary. Irrigation was done at 30  
 80 and 60 days after planting (DAP).

### 81 2.3 Data Collection

82 Leaves were collected at 30, 60, 90 and 120 DAP and chlorophyll a, chlorophyll b,  
 83 chlorophyll a/b ratio, total chlorophyll and carotenoids contents were estimated.

### 84 2.4 Sample Preparation and Chemical Analysis

85 Collected fresh leaves were cut into small pieces leaving away mid ribs, mixed thoroughly  
 86 and 250 mg of leaf materials were taken in a mortar and small amount of sodium carbonate  
 87 ( $\text{Na}_2\text{CO}_4$ ) was added into it to check the degradation of pigments. Leaf materials were  
 88 grinded finely by a pestle with 25 ml of cold 80% acetone for two minutes. Sample tubes  
 89 were centrifuged for 10 minutes (Model- SORVALL Legend Micro 21R Centrifuge, Thermo  
 90 Fisher Scientific, Germany). The homogenate was filtered through Whatman number 1 filter  
 91 paper and made up to 25 ml with cold 80% acetone. The centrifuged samples were  
 92 incubated in dark for half an hour. The optical density (OD) for each solution was measured  
 93 at 663, 645 and 440.5 nm (Model T80+, UV/VIS Spectrometer, PG Instruments Ltd., UK)  
 94 against 80% acetone as blank in one cm cell. The amount of chlorophyll-a and chlorophyll-b  
 95 were determined by using specific absorption coefficient of McKinney [19] and the formula of  
 96 Maclachalan and Zalik [20], Duxbury and Yentsch [21]. The amount of carotenoids was  
 97 determined by the equation of Holm [22].  
 98

99 i)  $C_a = \frac{(12.3 \times D_{663} - 0.86 \times D_{645})V}{d \times 1000 \times W}$  mg/g fresh leaf [20]

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 101 ii)  $C_b = \frac{(19.3 \times D_{645} - 3.6 \times D_{663})V}{d \times 1000 \times W}$  mg/g fresh leaf [21]

102  
 103 iii)  $C_c = 4.695 \times D_{440.5} - 0.268 C (a+b)$  [22]

104  
 105 iv) Total chlorophyll ( $\text{mg g fresh leaf}^{-1}$ ) = Chlorophyll a + Chlorophyll b

106  
 107 v) Chlorophyll-a and Chlorophyll-b ratio =  $\frac{\text{Chlorophyll a}}{\text{Chlorophyll b}}$

108 Where,

109 Ca = Chlorophyll a ( $\text{mg g fresh leaf}^{-1}$ )

110 Cb = Chlorophyll b ( $\text{mg g fresh leaf}^{-1}$ )

111 D = Optical density (O.D.) at wave length indicated

112 V = Final volume (ml)

113 W = Fresh weight of leaf materials used (g)

114 d = Length of light path (cm)

115 Cc = Concentration of carotenoids in  $\mu\text{g ml}^{-1}$ , which was then converted into mg  
 116  $\text{g}^{-1}$  fresh leaf

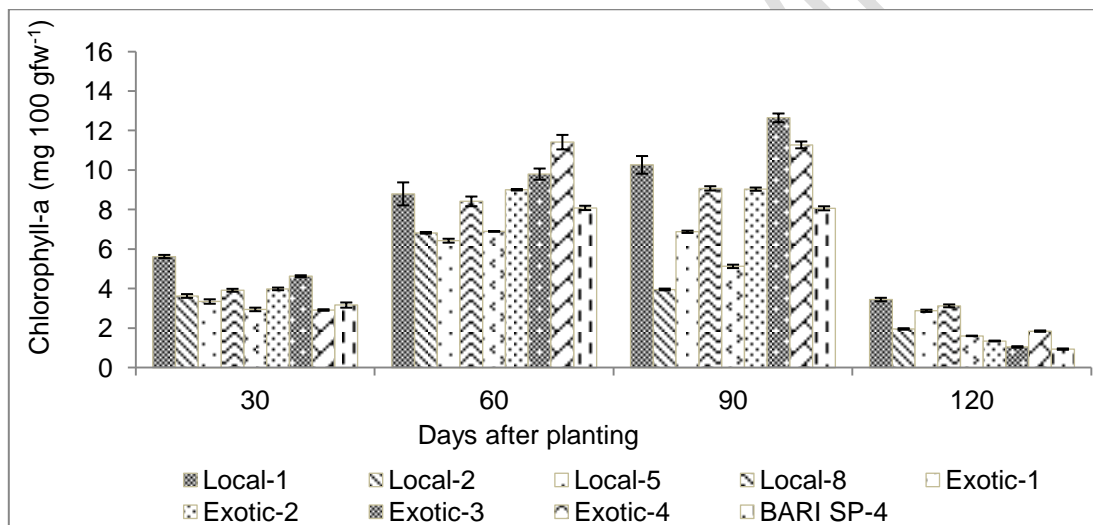
117 **2.5 Statistical Analysis of Data**

118 The data were analyzed using Analysis of Variance (ANOVA) technique through MSTATC  
119 package. Comparative analysis of the results was done using Duncan's Multiple Range Test  
120 (DMRT) at 1% level of significance. A p-value  $p \leq 0.01$  was considered statistically  
121 significant.  
122

123 **3. RESULTS AND DISCUSSION**

124 **3.1 Chlorophyll Content in Leaves**

125 Chlorophyll-a content ( $\text{mg } 100 \text{ gfw}^{-1}$ ) in leaves of all genotypes show that it gradually  
126 increased up to 60 DAP, after that it increased sharply in Exotic-4, Exotic-3 and Local-1, and  
127 decreased in Local-2, Local-5 and Exotic-1 up to 90 DAP (Fig. 1). After 90 days of planting,  
128 chlorophyll-a content decreased sharply up to 120 DAP. Local-1 produced the highest  
129 amount of chlorophyll-a ( $5.63 \pm 0.08$ ) at 30 DAP, increased to  $8.80 \pm 0.58$  at 60 DAP,  
130  $10.27 \pm 0.45$  at 90 DAP and finally reduced to  $3.45 \pm 0.09$  at 120 DAP.  
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**Fig. 1. Effect of genotype on the chlorophyll-a content in leaves of sweetpotato at different days after planting (DAP)**

**\*\*  $P \leq 0.01$ , Mean  $\pm$  S.E.M = Mean values  $\pm$  Standard error of means,  $n=3$ .**

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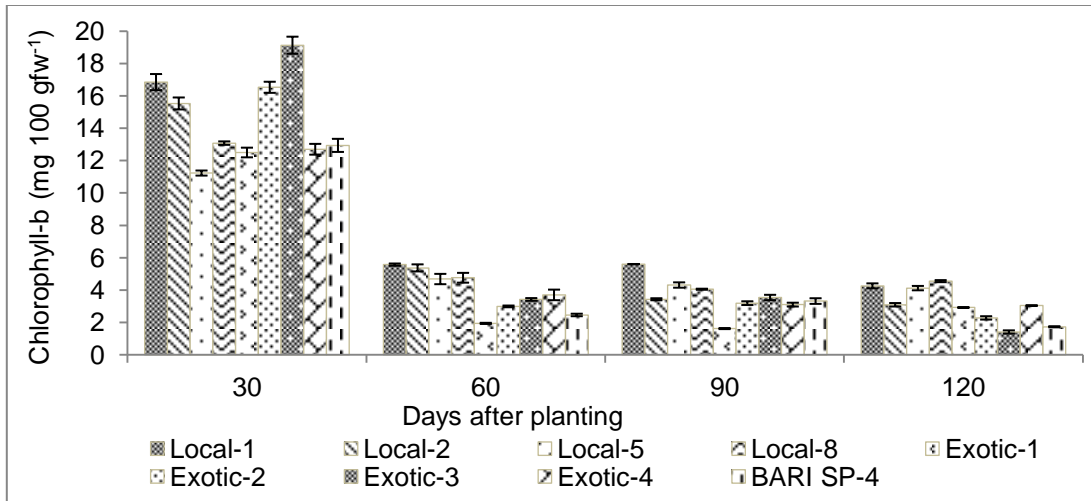
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Chlorophyll-b content in leaves ( $\text{in mg } 100 \text{ gfw}^{-1}$ ) had significant variations (Fig. 2). At initial stage, the amount of chlorophyll-b was higher in all of the genotypes and thereafter decreased dramatically. At 30 DAP, the highest amount of chlorophyll-b ( $\text{mg } 100 \text{ gfw}^{-1}$ ) was found in Exotic-3 ( $19.13 \pm 0.53$ ) followed by Local-1 ( $16.85 \pm 0.50$ ). At 60 DAP, the highest amount of chlorophyll-b was in Local-1 ( $5.57 \pm 0.07$ ) followed by Local-2 ( $5.37 \pm 0.22$ ) and Local-8 ( $4.76 \pm 0.30$ ) and the lowest was in Exotic-1 ( $1.94 \pm 0.04$ ). At 90 DAP, the highest amount was in Local-1 ( $5.60 \pm 0.01$ ) followed by Exotic-3 ( $4.54 \pm 0.17$ ) and the lowest was in Exotic-1 ( $1.62 \pm 0.03$ ). At 120 DAP, the highest amount was in Local-8 ( $4.55 \pm 0.06$ ) followed by Local-1 ( $4.26 \pm 0.15$ ) and the lowest was in Exotic-3 ( $1.41 \pm 0.10$ ). The ratio of chlorophyll-a to chlorophyll-b in higher plants is approximately 3:1 [7].

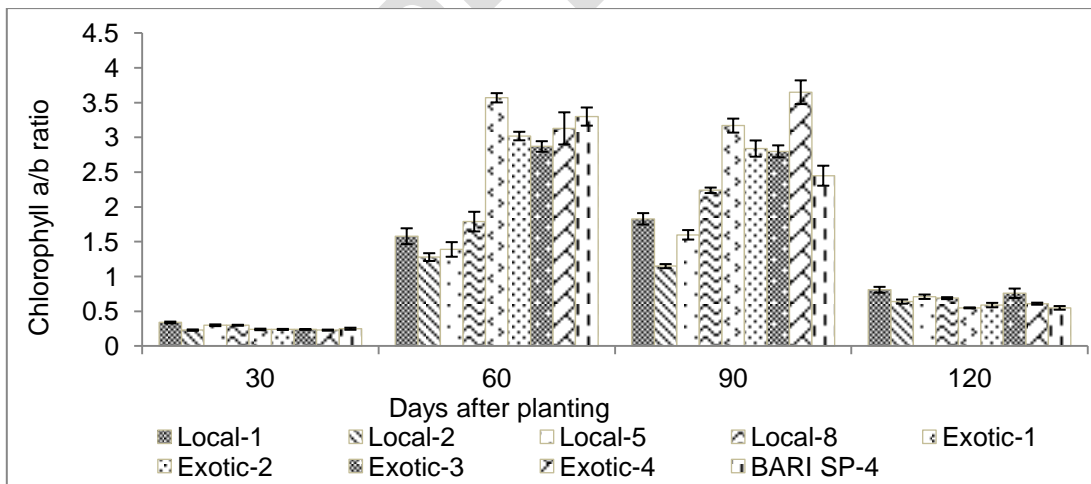


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**Fig. 2. Effect of genotype on the chlorophyll-b content in leaves of sweetpotato at different DAP**

**\*\*  $P \leq 0.01$ , Mean  $\pm$  S.E.M = Mean values  $\pm$  Standard error of means,  $n=3$ .**

151 As like as chlorophyll-a and chlorophyll-b, the ratio of them was also very low at initial stage  
152 and as days passes the ratio was increased up to 60 DAP (Fig. 3). From 60-90 DAP, ratio of  
153 Local-1, Local-5, Local-8 and Exotic-4 increased whereas ratio of Local-2, Exotic-1, Exotic-2,  
154 Exotic-3 and BARI SP-4 decreased gradually. After 90 days of planting, ratio decreased  
155 harshly. The highest ratio ( $3.65 \pm 0.17$ ) was in Exotic-4 and the lowest was in Local-2  
156 ( $1.15 \pm 0.03$ ) at 90 DAP. The chlorophyll a/b ratio of Local-1 was seen  $0.34 \pm 0.01$  at 30 DAP,  
157  $1.58 \pm 0.11$  at 60 DAP,  $1.81 \pm 0.08$  at 90 DAP and  $0.81 \pm 0.04$  at 120 DAP.  
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**Fig. 3. Effect of genotype on the chlorophyll a/b ratio in leaves of sweetpotato at different DAP**

**\*\*  $P \leq 0.01$ , Mean  $\pm$  S.E.M = Mean values  $\pm$  Standard error of means,  $n=3$ .**

164 Total chlorophyll consists of chlorophyll-a and chlorophyll-b varied significantly (Table 1).  
165 Initially although the total chlorophyll content ( $\text{mg } 100 \text{ gfw}^{-1}$ ) was high but it reduced with  
166 plant ages in all of the genotypes. After 30 days of planting, the highest total chlorophyll  
167 content was in Exotic-3 ( $23.76 \pm 0.56$ ) followed by Local-1 ( $22.48 \pm 0.46$ ) and the lowest in

168 Local-5 (14.58±0.32). After 60 days of planting, the highest total chlorophyll content was in  
 169 Exotic-4 (15.12±0.60) followed by Local-1 then Exotic-3 and Local-8. The lowest was in  
 170 Exotic-1 (8.84±0.04). After 90 days of planting, the highest total chlorophyll content was in  
 171 Exotic-3 (17.18±0.34) followed by Local-1 (15.87±0.44) and the lowest were in Exotic-1  
 172 (6.75±0.08). At final harvest, Local-1 (7.71±0.13) and Local-8 (7.68 mg) were showed similar  
 173 content of total chlorophyll.  
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 175

**Table 1. Total chlorophyll content in leaves of sweetpotato genotypes at different DAP**

Genotypes	Total chlorophyll content in leaves at different DAP (mg 100 gfw <sup>-1</sup> )			
	30	60	90	120
Local-1	22.48±0.46 b	14.37±0.55 ab	15.87±0.44 b	7.71±0.13 a
Local-2	19.16±0.33 d	12.20±0.20 cd	7.38±0.06 g	5.04±0.09 c
Local-5	14.58±0.14 g	11.11±0.28 de	11.19±0.15 f	7.00±0.12 b
Local-8	16.99±0.09 e	13.18±0.28 bc	13.13±0.10 d	7.68±0.12 a
Exotic-1	15.45±0.27 f	8.84±0.04 f	6.75±0.08 g	4.53±0.02 d
Exotic-2	20.52±0.32 c	12.00±0.05 cd	12.22±0.10 e	3.63±0.09 e
Exotic-3	23.76±0.56 a	13.22±0.3 bc	17.18±0.34 a	2.46±0.09 f
Exotic-4	15.62±0.32 f	15.12±0.60 a	14.38±0.15 c	4.89±0.03 c
BARI SP-4	16.10±0.36 f	10.54±0.10 e	11.39±0.15 ef	2.67±0.03 f
CV (%)	1.51	4.23	3.10	2.45
LSD <sub>.01</sub>	0.657	1.239	0.902	0.292

176 *Figures (Mean ± S.E.M) in a column having same letters do not differs significantly at 0.01*  
 177 *level of significane by DMRT*

178 The variations in chlorophyll-a, chlorophyll-b, chlorophyll and their corresponding a/b ratio  
 179 and total chlorophyll are probably due to genotypic, fertilization as well as growth stages.  
 180 The results corroborate with findings of Katayama and Shida [23] and Yooyongwech *et al.*  
 181 [24]. Katayama and Shida [23] reported chl a, chl b and ratio of a/b were 69.1 mg 100 gfw<sup>-1</sup>,  
 182 23.5 mg 100 gfw<sup>-1</sup> and 2.949, respectively in the leaves of 6th position in the sweetpotato  
 183 vine. They reported that the contents of chlorophyll a and b will change in their absolute  
 184 amount and also in their ratio a/b according to the kind of materials or to the different  
 185 developmental stages as well as fertilizers, chemicals, moisture and other environments.  
 186 They added that the change of chlorophyll contents was observed corresponding to the  
 187 developmental stages of leaves in sweetpotato. Rashid [25] established that the content of  
 188 chlorophyll-a, chlorophyll-b, and their ratio were influenced by the cultivar.  
 189

190 On the other hand, Yooyongwech *et al.* [24] reported the chlorophyll a, chlorophyll b, total  
 191 chlorophyll contents and ratio of Chl a : Chl b in three genotypes of sweetpotato grown under  
 192 well watering in the pot culture were ranged 24.07-33.46 mg 100 gfw<sup>-1</sup>, 11.75-14.43 mg 100  
 193 gfw<sup>-1</sup>, 36.95-47.89 mg 100 gfw<sup>-1</sup> and 1.71-2.32 mg 100 gfw<sup>-1</sup>. They reported that the

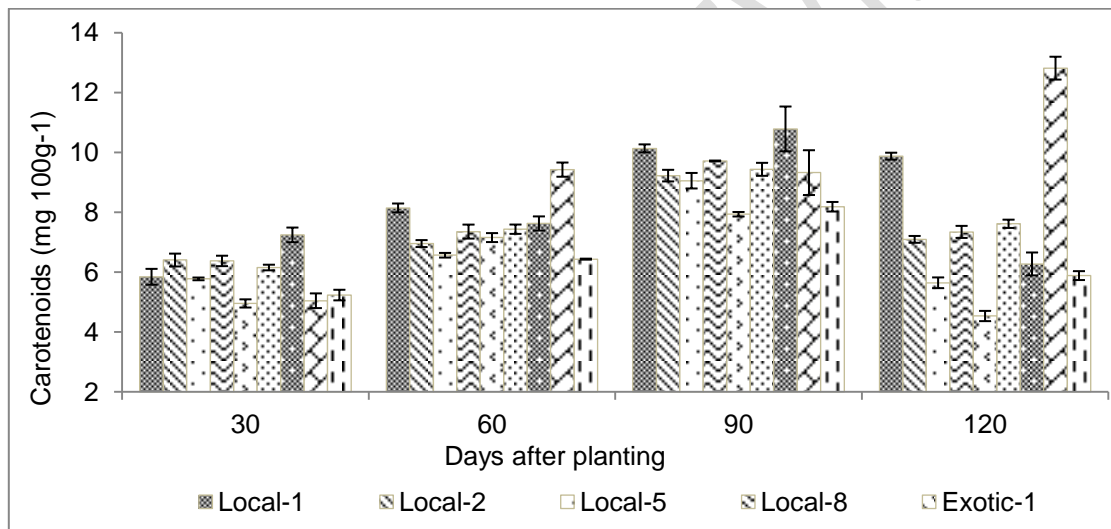
194 variation of the chlorophyll contents of the present result was perhaps due to genetic  
195 makeup.

### 196 3.2 Carotenoids Content in Leaves

197 Carotenoids content ( $\text{mg } 100 \text{ gfw}^{-1}$ ) of all genotypes increased gradually up to 90 DAP and  
198 thereafter decreased except Exotic-4 (Fig. 4). At 90 DAP, the highest carotenoids was in  
199 Exotic-3 (10.78 mg) followed by Local-1 (10.13 mg) and the lowest was in Exotic-1 (7.93  
200 mg). Overall, all of the genotypes were found better over the check variety BARI SP-4 in  
201 carotenoids production except Exotic-1.

202 The variations in carotenoids among the genotypes may be genotypic and/or environmental  
203 conditions. The results are in line of the works of the following researchers. Woolfe [4]  
204 reported carotenoids in sweetpotato leaves ranged from 0.38-7.24  $\text{mg } 100 \text{ gfw}^{-1}$ . Motsa *et al.*  
205 [26] reported that total carotenoids and total chlorophyll content of edible leaves were ranged  
206 from 90 to 390  $\text{mg } 100 \text{ gdw}^{-1}$  and 1.54 to 4.47  $\text{mg } 100 \text{ gdw}^{-1}$ , respectively. They added that  
207 total carotenoids and chlorophyll content in leaves were significantly affected by  
208 environmental conditions.

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212 **Fig. 4. Effect of genotype on the carotenoids content in leaves of sweetpotato at**  
213 **different DAP**

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\*\*  $P \leq 0.01$ , Mean  $\pm$  S.E.M = Mean values  $\pm$  Standard error of means,  $n=3$ .

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### 216 3.3 Total Dry Matter ( $\text{g plant}^{-1}$ )

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218 Total dry matter (TDM) increased gradually up to 60 DAP and thereafter increased very  
219 rapidly upto 120 DAP in all genotypes (Table 2). At 120 DAP, the highest TDM was in Local-  
220 8 ( $327.10 \pm 5.52$ ) followed by Local-1 ( $292.30 \pm 5.65$ ), and the lowest was in Exotic-3  
221 ( $116.90 \pm 1.36$ ).

222

223 Hossain and Islam [27] reported that total dry weights of 10 sweetpotato genotypes  
224 increased up to 165 DAP. Nandi and Sen [28] reported that the total biomass yield increased  
225 linearly upto 120 DAP except two genotypes. Nair and Nair [29] reported a linear increase in  
226 TDM. Mannan *et al.* [30] established that storage root DM increased rapidly from 90 to 150  
227 DAP. Haque [31] reported that TDM had a linear growth phase that continued until about

228 120 DAP. Watson [32] stated that total dry matter production of a crop is dependent on the  
 229 source and its activities as well as the length of its growth period, during which  
 230 photosynthesis continues. The above results agree with the present result.

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**Table 2. Effect of genotypes on the total dry matter of sweetpotato at different DAP**

Genotypes	Total dry matter (g plant <sup>-1</sup> )			
	30	60	90	120
Local-1	23.03±1.22 a	34.87±1.16 a	100.30±1.22 b	292.30±5.65 b
Local-2	17.97±0.50 b	32.38±0.43 b	93.82±0.96 c	232.90±2.39 c
Local-5	12.08±0.46 c	23.42±0.52 d	74.07±1.64 e	170.90±2.87 e
Local-8	15.32±0.28 b	32.06±0.31 b	118.70±2.97 a	327.10±5.52 a
Exotic-1	15.28±0.49 b	22.72±0.43 d	56.85±0.71 fg	134.20±1.57 g
Exotic-2	16.27±0.87 b	28.19±0.70 c	77.56±1.83 e	157.10±0.56 f
Exotic-3	9.11±0.22 d	16.98±0.50 e	51.68±1.66 g	116.90±1.36 h
Exotic-4	16.53±0.52 b	24.66±0.22 d	62.76±1.15 f	175.80±1.50 e
BARI SP-4	11.80±0.09 c	23.99±0.33 d	85.46±1.31 d	209.10±3.13 d
CV%	7.20	3.89	3.32	2.69
Lsd .01	2.622	2.468	6.350	12.970

234 Figures (Mean ± SEM) in a column having similar letters do not differ significantly at 1% level  
 235 of significance by DMRT

### 236 3.4 Storage Root Dry Weight (g plant<sup>-1</sup>)

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238 Storage roots dry weight increased gradually up to 90 DAP and then increased sharply up to  
 239 120 DAP (Table 3). After 30 days of planting, storage roots appeared only in Local-8  
 240 (0.65±0.09) and check variety BARI SP-4 (1.88±0.08). After 60 days of planting, all of the  
 241 genotypes initiated storage roots in except Exotic-1 and Exotic-4. After 90 days of planting,  
 242 all of the genotypes initiated storage roots. After 120 days of planting, the highest weight  
 243 was in Local-8 (232.40±5.97), followed by Local-1 (187.50±5.23) and the lowest weight was  
 244 in Exotic-1 (54.05±1.11).

245

246 The above results corroborate with the findings of Oswald *et al.* [33] where the storage root  
 247 dry matter of sweetpotato increment followed a sigmoid pattern, and Nair and Nair [29] while  
 248 they reported a linear increase in storage root dry matter and the increase in storage root dry  
 249 matter was the maximum during 48 to 161 days of planting.

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253 **Table 3. Effect of genotypes on the storage roots dry weight of sweetpotato at**  
 254 **different DAP**

Genotypes	Storage root dry weight (g plant <sup>-1</sup> )
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	30	60	90	120
Local-1	0.00±0.00 c	2.73±0.04 de	31.13±0.46 c	187.50±5.23 b
Local-2	0.00±0.00 c	4.30±0.14 c	29.88±0.80 cd	152.90±1.61 c
Local-5	0.00±0.00 c	2.90±0.05 d	25.08±1.36 de	98.28±1.53 d
Local-8	0.65±0.09 b	7.97±0.02 b	60.09±2.47 a	232.40±5.97 a
Exotic-1	0.00±0.00 c	0.00±0.00 g	13.07±0.91 f	54.05±1.11 f
Exotic-2	0.00±0.00 c	2.57±0.09 e	29.18±0.91 cd	86.42±1.31 d
Exotic-3	0.00±0.00 c	1.66±0.06 f	22.80±1.62 e	68.98±0.32 e
Exotic-4	0.00±0.00 c	0.00±0.00 g	8.720±0.15 f	93.39±1.55 d
BARI SP-4	1.88±0.08 a	8.27±0.08 a	53.24±0.22 b	144.20±3.14 c
CV%	17.48	3.48	7.37	4.12
Lsd .01	0.1067	0.2822	5.337	12.22

255 Figures (Mean ± SEM) in a column having similar letters do not differ significantly at 1% level  
256 of significance by DMRT

### 257 3.5 Correlation of Chlorophyll and Carotenoids with Yield of Sweetpotato

258 At 30 DAP chlorophylls and carotenoids had no significant effect on total dry matter and  
259 storage roots dry weight (Table 4). However, Chlorophyll-a was highly correlated with  
260 chlorophyll-b, total chlorophyll, chlorophyll a /b ratio and carotenoids whereas chlorophyll-b  
261 with total chlorophyll and carotenoids.

262 After 60 days of planting chlorophyll-b had positive significant correlation with TDM while  
263 chlorophyll a/b ratio correlated negatively (Table 5). Carotenoids had negative correlation  
264 with storage roots dry weight and had positive correlation with chlorophyll-a and total  
265 chlorophylls. Same correlation was observed at 90 DAP except carotenoids with storage root  
266 dry weight (Table 6).

267 After 120 days of planting it is seen that all chlorophyll components and carotenoids had  
268 positive correlation with TDM and storage roots dry weight (Table 7) while chlorophyll-a,  
269 chlorophyll-b and total chlorophylls had significant correlation with them. It is also indicates  
270 that Chlorophyll-a had no effect on storage roots dry weight upto 90 DAP while chlorophyll-b  
271 and total chlorophyll upto 60 DAP. Chlorophyll-b had positive correlation with TDM from the  
272 beginning while chlorophyll-a, total chlorophyll and chlorophyll a/b ratio was negative or  
273 insignificant upto 120 DAP.

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275

276 **Table 4. Correlation of chlorophyll and carotenoids with total dry matter and storage**  
277 **root dry weight (g plant<sup>-1</sup>) at 30 DAP**

TDM	SDW	Chl-a	Chl-b	Total chl	Chl a/b ratio
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SDW	-0.307					
Chl-a	0.375	-0.250				
Chl-b	0.099	-0.282	0.722**			
Total chl	0.178	-0.290	0.836**	0.983**		
Chl a/b ratio	0.333	-0.049	0.612**	-0.096	0.086	
Carotenoids	-0.187	-0.218	0.514**	0.731**	0.715**	-0.011

278 TD = Total dry matter (g plant<sup>-1</sup>), SDW = Storage root dry weight (g plant<sup>-1</sup>), Chl-a = Chlorophyll-a, Chl-  
279 b = Chlorophyll-b, Chl a/b ratio = Ratio chlorophyll-a and chlorophyll-b

280 **Table 5. Correlation of chlorophyll and carotenoids with total dry matter and storage**  
281 **root weight (g plant<sup>-1</sup>) at 60 DAP**

	TDM	SDW	Chl-a	Chl-b	Total chl	Chl a/b ratio
SDW	0.349					
Chl-a	-0.152	-0.260				
Chl-b	0.645**	0.179	-0.093			
Total chl	0.301	-0.094	0.754**	0.583**		
Chl a/b ratio	-0.587**	-0.275	0.396*	-0.936**	-0.295	
Carotenoids	0.089	-0.522**	0.775**	0.138	0.723**	0.165

282 TD = Total dry matter (g plant<sup>-1</sup>), SDW = Storage root dry weight (g plant<sup>-1</sup>), Chl-a = Chlorophyll-a, Chl-  
283 b = Chlorophyll-b, Chl a/b ratio = Ratio chlorophyll-a and chlorophyll-b

284 **Table 6. Correlation of chlorophyll and carotenoids with total dry matter and storage**  
285 **root weight (g plant<sup>-1</sup>) at 90 DAP**

	TDM	SDW	Chl-a	Chl-b	Total chl	Chl a/b ratio
SDW	0.783**					
Chl-a	-0.171	-0.037				
Chl-b	0.397*	0.282	0.487**			
Total chl	-0.010	0.061	0.960**	0.712**		
Chl a/b ratio	-0.571**	-0.391*	0.478*	-0.510**	0.221	
Carotenoids	0.088	0.004	0.610**	0.713**	0.719**	-0.165

286 TD = Total dry matter (g plant<sup>-1</sup>), SDW = Storage root dry weight (g plant<sup>-1</sup>), Chl-a = Chlorophyll-a, Chl-  
287 b = Chlorophyll-b, Chl a/b ratio = Ratio chlorophyll-a and chlorophyll-b

288 **Table 7. Correlation of chlorophyll and carotenoids with total dry matter and storage**  
289 **root weight (g plant<sup>-1</sup>) at 120 DAP**  
290

	TDM	SDW	Chl-a	Chl-b	Total chl	Chl a/b ratio
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SDW	0.984**					
Chl-a	0.715**	0.634**				
Chl-b	0.686**	0.591**	0.950**			
Total chl	0.708**	0.618**	0.985**	0.990**		
Chl a/b ratio	0.303	0.306	0.563**	0.301	0.425*	
Carotenoids	0.268	0.202	0.252	0.230	0.243	0.148

291 TD = Total dry matter (g plant<sup>-1</sup>), SDW = Storage root dry weight (g plant<sup>-1</sup>), Chl-a =  
 292 Chlorophyll-a, Chl-b = Chlorophyll-b, Chl a/b ratio = Ratio chlorophyll-a and chlorophyll-b

### 293 **3.4 Contribution to the Existing Knowledge/Implication of the Study**

294 Piedmont soil is acidic in nature. Sweetpotato is capable of rapid coverage, so research on  
 295 acid reclamation and reducing soil erosion through cultivation of Local-1, Local-5, Local-8  
 296 and Exotic-1 need to be initiated. These genotypes may be incorporated in Jum cultivation in  
 297 hilly areas.

### 298 **4. CONCLUSION**

300 Chlorophyll-a content decreased after 60 days of planting while chlorophyll-b decreased  
 301 from 30 DAP and carotenoids decreased from 90 DAP. At 120 DAP, the higher chlorophyll  
 302 and carotenoids synthesizing genotypes were Local-1, Local-8, Exotic-3 and Exotic-4, and  
 303 storage roots dry weight obtaining genotypes were Local-8 and Local-1. Chlorophyll-b had  
 304 positive correlation with TDM from the beginning while chlorophyll-a, total chlorophyll and  
 305 chlorophyll a/b ratio had negative or insignificant effect on TDM up to 120 DAP. Chlorophyll-  
 306 a had positive effect on storage roots dry weight after 90 DAP while chlorophyll-b and total  
 307 chlorophyll came to positive effect on storage roots dry weights after 60 DAP.

### 310 **COMPETING INTERESTS**

311 Authors have declared that no competing interests exist.

### 312 **REFERENCES**

- 313 [1] Ministry of Agriculture, Govt. of Bangladesh. Crop situation in Bangladesh. 2017.  
 314 Available: [www.moa.gov.bd](http://www.moa.gov.bd), [http://F:/sweetpotato/information/crop\\_situation\\_2014-16.pdf](http://F:/sweetpotato/information/crop_situation_2014-16.pdf).  
 315
- 316 [2] Food and Agriculture Organization. FAO Statistical Year Book of the United Nations,  
 317 Regional Office for Asia and the Pacific Bangkok. 2014; 176. Available at: [www.fao.org](http://www.fao.org)  
 318
- 319 [3] Kozai T, Kubota C and Kitaya Y. Sweetpotato technology for solving the global issues on  
 320 food, energy, natural resources and environment in the 21st century. Environ. Control in Biol.  
 321 2006; 34:105–114.  
 322
- 323 [4] Woolfe JA. Sweetpotato: An untapped food resource, Cambridge University Press,  
 324 Cambridge, UK. 1992; pp. 643.  
 325  
 326  
 327  
 328  
 329

- 330  
331 [5] Laurie SM, Faber M, van Jaarsveld PJ, Laurie RN, du Plooy CP, Modisane PC. Beta-  
332 carotene yield and productivity of orange-fleshed sweetpotato (*Ipomoea batatas* (L.) Lam.)  
333 as influenced by irrigation and fertilizer application treatments. *Sci. Hortic.* 2012; 142:180-  
334 184.
- 335  
336 [6] Islam S. Nutritional and medicinal qualities of sweetpotato tops and leaves. Cooperative  
337 Extension Program, University of Arkansas, Pine Bluff, Chicago, USA. 2014.
- 338  
339 [7] Arnon DI. Copper enzyme in isolated chloroplast polyphenol oxidase in *Beta vulgaris* (L.).  
340 *Plant Physiology.* 1949; 24:1-5.
- 341  
342 [8] Abramavicius D, Valkunas L. Role of coherent vibrations in energy transfer and  
343 conversion in photosynthetic pigment-protein complexes. *Photosynth. Res.* 2016; 127, 33-  
344 47.
- 345  
346 [9] Batjuka A, Skute N, Petjukevics A. The influence of anthocyanin on pigment composition  
347 and functional activity of photosynthetic apparatus of *Triticum aestivum* L. under high  
348 temperature. *Photosynthetica.* 2016; 55: 1–14.
- 349  
350 [10] Zakar T, Laczko-Dobos H, Toth TN, Gombos Z. Carotenoids assist in cyanobacterial  
351 photosystem II assembly and function. *Front. Plant Sci.* 2016; 7:295.
- 352  
353 [11] Campos MD, Nogales A, Cardoso HG, Campos C, Grzebelus D, Velada I *et al.* Carrot  
354 plastid terminal oxidase gene (dcptox) responds early to chilling and harbors intronic pre-  
355 miRNAs related to plant disease defense. *Plant Gene.* 2016; 7:21–25.
- 356  
357 [12] Santabarbara S, Casazza AP, Ali K, Economou CK, Wannathong T, Zito F *et al.* The  
358 requirement for carotenoids in the assembly and function of the photosynthetic complexes in  
359 *Chlamydomonas reinhardtii*. *Plant Physiol.* 2013; 161:535–546.
- 360  
361 [13] Nagy L, Kiss V, Brumfeld V, Osvay K, Borzsonyi A, Magyar M *et al.* Thermal effects and  
362 structural changes of photosynthetic reaction centers characterized by wide frequency band  
363 hydrophone: Effects of carotenoids and terbutryn. *Photochem. Photobiol.* 2015; 91:1368–  
364 1375
- 365  
366 [14] Kopsell DA, Kopsell DE, Curran-Celentano J Carotenoid and chlorophyll pigments in  
367 sweet basil grown in the field and greenhouse. *HortScience.* 2005; 40 (5):1230–1233.
- 368  
369 [15] Rumbaoa RG, Cornago DF and Geronimo IM Phenolic content and antioxidant capacity  
370 of Philippine sweetpotato (*Ipomoea batatas*) varieties. *Food Chem.* 2009; 113:1133–1138.
- 371  
372 [16] Hue SM, Boyce AN, Chandran S. Influence of growth stage and variety on the pigment  
373 levels in *Ipomoea batatas* (sweetpotato) leaves. *African Journal of Agricultural Research.*  
374 2011; 6(10):2379-2385.
- 375  
376 [17] Rajput A, Rajput SS, Jha G. Physiological parameters leaf area index, crop growth rate,  
377 relative growth rate and net assimilation rate of different varieties of rice grown under  
378 different planting geometries and depths in SRI. *Int. J. Pure App. Biosci.* 2017; 5(1):362-367.  
379 doi: <http://dx.doi.org/10.18782/2320-7051.2472>
- 380  
381 [18] SRDI (Soil Resource Development Institute). Land and soil statistical appraisal book of  
382 Bangladesh, 1st edn., Ministry of Agriculture, Govt. of Bangladesh. Dhaka. 2010; pp.11-103.

383  
384 [19] Mackinney G. Criteria for purity of chlorophyll preparations. *Journal of Biological*  
385 *Chemistry*. 1940; 132:91-109.  
386  
387 [20] Maclachlan S, Zalik S. Plastid structure, chlorophyll concentration, and free amino acid  
388 composition of a chlorophyll mutant of barley. *Canadian Journal of Botany*. 1963; 41: 1053-  
389 1062.  
390  
391 [21] Duxbury AC, Yentsch CS. Plant and pigment monography. *Journal of Air Pollution and*  
392 *Control Assessment*. 1956; 16:145-150.  
393  
394 [22] Holm G. Chlorophyll mutations in Barley. *Acta Agric. (Scand.)*. 1954; 4:457-461.  
395  
396 [23] Katayama Y, Shida S. Studies on the change of chlorophyll a and b contents due to  
397 projected materials and some environmental conditions. *Cytologia*. 1970; 35:171-180.  
398  
399 [24] Yooyongwech S, Samphumphuang T, Theerawitaya C, Chaum S. Physio-morphological  
400 responses of sweetpotato [*Ipomoea batatas* (L.) Lam.] genotypes to water-deficit stress. *J.*  
401 *Plant Omics*. 2014; 7(5):361-368.  
402  
403 [25] Rashid MMHA. Comparative growth and yield studies of some exotic and local cultivars  
404 of sweetpotato. M.Sc. (Agriculture) Thesis, Dept. of Crop Botany, Bangladesh Agricultural  
405 University, Mymensingh. 2002.  
406  
407 [26] Motsa NM, Modi AT, Mabhaudhi T. Influence of agro-ecological production areas on  
408 antioxidant activity, reducing sugar content, and selected phytonutrients of orange-fleshed  
409 sweetpotatocultivars. *Food Sci. Technology (Cam-606ea batatas pinas)*. 2015; 35(1):32-37.  
410 [27] Hossain MAS, Islam AFMS. Comparative study on dry matter partitioning in five exotic  
411 genotypes and five local varieties of sweetpotato. *J. Sher-e-Bangla Agric. University*. 2010;  
412 4(1):24-30.  
413  
414 [28] Nandi S, Sen H. Evaluation of sweetpotato (*Ipomoea batatas* L. Lam.) cultivars under  
415 late planted situation. *Root Crops J*. 1998; 24(1):73-77.  
416  
417 [29] Nair GM, Nair VM. Influence of irrigations and fertilizers on the growth attributes of  
418 sweetpotato, *Journal of Root Crops*. 1995; 21:17-23.  
419  
420 [30] Mannan MA, Bhuiyan MKR, Quasem A, Rashid MM, Siddique MA. Study on the  
421 growth and partitioning of dry matter in sweet potato. *J. Root Crops*. 1992; 18:1-5.  
422  
423 [31] Haque MM. Productivity of Maize/sweetpotato intercropping in relation to planting  
424 system, population density and application of nitrogen and potassium fertilizers, PhD  
425 Dissertation, Dept. of Agronomy, Institute of Post-Graduate Studies in Agriculture, Salna,  
426 Gazipur-1703, Bangladesh.1995; pp.164.  
427  
428 [32] Watson DJ. The dependence of net assimilation rate on leaf area index. *Ann. Bot.* 1958;  
429 22:37-54.  
430  
431 [33] Oswald A, Alkamper J, Midmore DJ. The effect of different shade levels on growth and  
432 tuber yield of sweetpotato, 1. Plant development. *Journal Agron. Crop Sci*. 1994; 175:99-  
433 107.