

**THE ANIMAL MODELS FOR LEUKAEMIA
INDUCTION USING CHEMICAL CARCINOGENS**

ABSTRACT

Animal models have been providing invaluable contributions to the better understanding of mechanisms of cancer (including leukaemias) development and effectiveness of most of the treatments. Chemical carcinogens are generally used to study the biology of cancers including leukaemias in many animal models, including rats and mice. The studies in most cases are aimed at development and evaluation of cancer treatments and preventions. Some of the most common chemical carcinogens used in animal models for leukaemias include *N*-ethyl-*N*-nitrosourea (ENU), *N*-methyl-*N*-nitrosourea (MNU), dimethyl benz(a)nthracene (DMBA) and benzo(a)pyrene (BaP). This review provides highlights on different animal models of leukaemia induced by the chemical carcinogens mentioned earlier, at the same time discussed the contributions of these models to the leukaemia diagnosis in laboratory animal models for subsequent development of treatment.

Keywords: Animal model, dimethyl benzanthracene (DMBA), Benzo (a) pyrene (BaP), Leukaemia, N-ethyl-N-nitrosourea (ENU), N-methyl-N-nitrosourea (MNU).

1 INTRODUCTION

Leukaemia refers to neoplastic proliferation of lymphoid and myeloid progenitor cells because of mutation of a single stem cell, the progeny of which form a clone of leukaemic cells. It is broadly classified into acute and chronic leukaemias and each is further subdivided into myeloid and lymphoid; i.e. acute myeloid leukaemia, acute lymphocytic leukaemia, chronic myeloid leukaemia, and chronic lymphocytic leukaemia [1]. It has been reported that, CLL is the most common type of leukaemia found, however, AML accounts for about 42% of all leukaemia deaths [2].

The causes of leukaemia are not well understood; however, alkylating drugs, ionizing radiation, and chemicals have been incriminated, these have been shown to induce chromosomal abnormalities resulting to DNA changes. [3] reported leukaemia to be the most common type of cancer developed following treatment with alkylating agents, predominantly nonlymphoid leukaemia.

Various animal models have been developed to investigate the factors involved in malignant transformation, invasion and metastasis, at the same time to investigate the effectiveness of the treatment (therapy). The importance of animal models on cancer research; leukaemia inclusive cannot be overemphasized. These includes studies on oncogenesis, molecular genetics, microenvironment, metastasis, therapeutic effects, etc. [4]. Biochemical researches are primarily designed to provide advanced knowledge which could be used to predict disease pathology and pathophysiology by a clinician and at the same time to choose appropriate treatment. Animal models can be used to test relationships and mechanisms under controlled experimental conditions which can be used to predict clinical outcomes in humans [4]. Animal models can also assist in identifying the exogenous agents to which the exposure may underlie leukaemic induction. Chemically induced leukaemic animal models

38 are generally used to study the pathogenesis of leukaemia to develop the treatment and
39 prevention of the cancer [5].

40 This review focused on chemical carcinogens used in different animal models to understand
41 the mechanisms of cancer development and the effectiveness of some of the treatments
42 tested.

43 The most commonly used chemical carcinogens in studying the progression of leukaemia
44 are *N*-ethyl-*N*-nitrosourea (ENU), *N*-methyl-*N*-nitrosourea (MNU), dimethyl benzanthracene
45 (DMBA), benzo (a) pyrene (BaP), amongst others. This review will focus on these chemicals
46 as they are commonly used in rat and mouse models.
47

48 **2 METHODOLOGY**

49 **2.1 Nitrosoureas**

50 Nitrosoureas (ENU, MNU, etc.) were extensively used in the treatment of cancers in the
51 past. They spontaneously decompose to generate two reactive species; namely an
52 alkylating group and a carbamoylating group, either of these may react with DNA, RNA or
53 protein, thereby causing serious and often prolonged bone marrow suppression [6]. The
54 mechanism of action of nitrosoureas involved transfer of its alkyl (ethyl or methyl) group to
55 the nucleobases of the cancer cells, this causes DNA changes. The transfer of the alkyl
56 group require the action of an enzyme; alkyl transferase from the bone marrow, this causes
57 decreased concentration of the enzymes within the bone marrow, leading to
58 leukaemogenesis [7]. Alkylation of DNA occurs relatively homogeneously throughout the
59 body due to the non-enzymatic formation of the reaction product. This is like most
60 chemotherapy drugs. which generally cause DNA damage without cytochrome p450-
61 mediated bioactivation. If a methyl or ethyl adduct is not removed by alkylguanine
62 transferase, the whole nuclear base adduct may be removed via excision repair. Removal of
63 the modified base and neighbouring nucleotides creates a strand break, which causes the
64 activation of the nuclear enzyme, poly(ADP-ribose) polymerase (PARP). Activated PARP
65 make use of NAD⁺ as a substrate in the synthesis of poly(ADP-ribose). It is usually the main
66 acceptor protein, however a number of other nuclear proteins became modified to some
67 extent [8]. The concentration of negatively charged poly(ADP-ribose) at the site of DNA
68 damage may play several roles, including regulation of excision repair, p53 function and
69 apoptosis [9]. The most consistent evidence, however, shows that poly(ADP-ribose)
70 synthesis prevents recombination events at the site of damage, perhaps by repelling other
71 DNA strands, thereby decreasing the risk of chromosomal translocations. This is critical in
72 the bone marrow because majority of leukaemias resulted from translocation events of this
73 nature [9].

74 **2.1.1 Leukaemia induced by N-ethyl-N-nitrosourea (ENU)**

75 Ethyl nitrosourea is a monofunctional ethylating agent that has been used as a simple model
76 of chemotherapeutic nitrosoureas. Monofunctional nitrosoureas are potent leukaemogens
77 that model the leukaemogenic aspect of chemotherapeutic agents quite well, especially
78 when used in certain strains of rats such as Long Evans, which respond with primarily
79 nonlymphocytic leukaemias. The leukaemic effects of ENU has been reported in both mice
80 and rats. A leukaemic induction protocol using intravenous injection in rats was previously
81 reported. Two different groups of rats; A and B were given IV injection of 225 mg/kg total
82 dose of ENU. Group A, comprising of 200 3-months old male Wistar rats, were administered
83 15 mg/kg ENU weekly for 15 weeks intravenously while group B comprising the same
84 number of animals at the same age received the same total dose of 225 mg/kg ENU by IV
85 injection of 75 mg/kg weekly for three weeks. After the last dose of ENU in each of the
86 groups, liver and spleens were palpated on weekly basis; any rats with splenomegaly and /or
87 hepatomegaly were subjected further to complete blood count, peripheral blood smears and
88 later liver biopsy to confirm the presence of leukaemia [10].

89 In group A leukaemia was observed in 91(45.5%) of the total animals; among these 76
90 (83.5%) were acute leukaemias diagnosed at 37 weeks whereas 15 (16.4%) were chronic
91 myeloid leukaemias which occurred at 30 weeks. However, group B on the other hand,
92 presented 63 (31.5%) total leukaemias, where 57 (90.5%) were acute with minimum
93 manifestation time of 35 weeks and 6 (9.5%) were chronic myeloid leukaemias with
94 minimum manifestation of 33 weeks [10].

95 The leukaemia presented in this research is characterized in addition to enlarged spleen,
96 liver and lymph nodes, by various sizes and shapes of blast cells from the peripheral blood.
97 The cells had round or oval nuclei with occasional indentations containing 1-3 nucleoli.
98 Moreover, the cytoplasm of the cells had no azurophilic or neutrophilic granules [10].

99 Different types of leukaemia, predominantly lymphoblastic was reported in albino mice (5-10
100 days old) injected with ENU at dose rate of 80 mg/kg intraperitoneally (IP) within 4-7 months
101 post inoculation [11]. Out of the total experimental animals, 15 % died due to acute toxicity
102 within 2-3 days after injection, 25% died from secondary infections and 60% presented a
103 progressive leukaemia status, characterized by leukaemic blasts cells in both the bone
104 marrow and peripheral blood smears [11].

105 The effect of IP administration of 80 mg/kg twice at one-week interval in 7-10 days old BALB
106 c male mice was also studied. Leukaemia was confirmed 5 months after the last ENU
107 injection by the appearance of numerous blast cells in peripheral blood and bone marrow
108 smears. Leucocytosis was also observed in the challenged group compared to the untreated
109 animal. However 2 of the 12 experimental animals died acutely (Bhattacharjee *et al.*, 2015).

110 Two years later, (Singha *et al.*, 2017) studied the effect of IP injection of ENU on older mice.
111 Three weeks old BALB/c mice were administered 80 mg/kg ENU IP twice at one-week
112 interval. In this case, five months post injection of the chemical, leukaemia was established,
113 characterized by the appearance of undifferentiated blast cells in the blood and bone marrow
114 smears, however no mortality was reported [13]. Perhaps due to the age of the experimental
115 animals in the study.

116 **2.1.2 Leukaemia induced by N-methyl-N-nitrosourea (MNU)**

117 The carcinogenic potential of MNU in Wistar rat was reported previously. It was
118 demonstrated that MNU promptly promotes malignant lymphoma and leukaemia [14]. Three
119 weeks old male Wistar rats weighing 220–230g were grouped into 4 groups; group A served
120 as the control, while groups B, C and D were respectively administered 20 mg/kg of body
121 weight (total of 80 mg/kg), 40 mg/kg of body weight (total of 160 mg/kg) and 60 mg/kg of
122 body weight (total of 240 mg/kg) MNU, intraperitoneally. A total of four injections (two
123 injections per week) were administered during the first two weeks of the experiments. The
124 animals were then closely observed and sacrificed at the 12th and 20th weeks, where
125 specific organs including thymus, spleen, bone marrow, cervical and mesenteric lymph
126 nodes and liver were collected for analysis. At the 20th week, lymphohematopoietic system
127 (LHS) malignant tumours and benign vascular tumours occurred only in the high- and
128 intermediate-dose MNU-treated animals. Four animals treated with 240 mg/kg developed
129 diffuse thymic lymphomas; two others, treated respectively with 240 mg/kg and 160 mg/kg,
130 developed spleen haemangiomas. Animals in group C demonstrated significant decreased
131 in the mean body weights, at the same time developed tumour of the thymus and spleen at
132 week 20 of the experiment [14]

133
134 Hutheya [15] studied the carcinogenic effect of MNU in Sprague Dawley rats where he
135 reported that IP injection of MNU twice weekly for two consecutive weeks at 60 mg/kg (total
136 dose of 240 mg/kg body weight) induced stage IV lymphoma characterized by enlargement
137 of the lymph nodes, hepatosplenomegaly (in 30% of the treated animals) and enlarged
138 kidneys in 7% of the rats. It was further reported that the organs were infiltrated with
139 malignant lymphocytes in different grades [15].

140 The effect of intravenous injection of MNU in Sprague Dawley rats has also been reported.
141 Seven weeks old SD rats were given a series of six IV injections of MNU at 35 mg/kg body

142 weight biweekly via the caudal vein and observed for a period of 220 days [1]. During the
143 period of observation, the animals were individually monitored daily for mortality and weekly
144 for clinical signs and body weight changes. Blood samples were analysed at 30-40 days
145 intervals after the final dose of MNU for complete blood count with differentials, cytology
146 (smears were prepared for Liu's and Papanicolaou stains) and serum biochemistry. The
147 animals were sacrificed humanely after 220th day of the experiment. Biopsies of liver and
148 spleen were prepared for both histopathological and immunohistochemical evaluations.
149

150 Leukaemia was observed in 97.1 % of the treated animals. The treated (leukaemic) group
151 showed less body weight compared to the untreated group with a significant increase in liver
152 and spleen weights. The treated rats also presented a significant leucocytosis with
153 neutropenia, however, the red blood cells (RBC) counts and haemoglobin (Hb) concentration
154 were decreased (which signifies tendency to anaemia). The study suggested that MNU
155 caused leucocytosis without affecting the neutrophil index. The increased production of blast
156 cells led to the reduction in the percentage of normal neutrophils and implied direct
157 correlation between leukaemia blasts and the number of white blood cells (WBCs) [1]. The
158 study reported further that, MNU was able to induce a significant increase in the serum
159 concentration of GOT and GPT, at the same time the uric acid level in the serum of
160 leukaemic rats was found to be higher than the control group. Cytologically, Papanicolaou
161 and Liu's staining techniques were employed for the analysis of the slides. More atypical
162 cells were found in the MNU treated group compared to the control. Papanicolaou staining
163 method, revealed cells that were round, dark blue in colour, and about two to three times the
164 size of lymphocytes. The Liu's stained smears in the MNU treated rats were cellular and
165 revealed markedly polymorphous mixture of small to large cells. It further revealed a larger
166 proportion of large, dark staining, blast-like promyelocytes and metamyelocytes as well as
167 dysplastic granulocytes [1]. Liu's staining method is considered more sensitive in the
168 classification of leukaemia/lymphoma, this is because it differentiates the lymphoblast from
169 myeloblasts more precisely than the Papanicolaou stain [1]. Histologically, there was
170 frequent enlargement of liver and spleen in all cases, which also presented leukaemic
171 cellular infiltration on the portal area and sinusoids of the liver. The normal architecture of
172 red and white pulp of the spleen were destroyed in MNU treated rats with massive spread of
173 undifferentiated cells. The immunohistochemical analysis revealed that CD3 (which
174 generated an active signal in T lymphocytes) and CD20 (which is active in all B-cell
175 lymphomas and leukaemia) expression were negative in malignant cells. However, MPO
176 (myeloperoxidase) staining was strongly positive in neoplastic cells, indicating that MNU
177 induced blast cells were derived from the myeloid lineage. This is because MPO stain
178 distinguishes myeloblastic from lymphoblastic leukaemias [16].
179

180 **2.2 Leukaemia induced by DMBA (7, 12 dimethylbenz[A]anthracene)**

181 Dimethylbenz[a]anthracene) is an immunosuppressor and considered a powerful organ
182 specific laboratory carcinogen that is widely used in many cancer researches as a tumour
183 initiator. Leukaemia due to DMBA was studied previously [17], by injecting pulse doses of
184 45-25 mg/kg body weight DMBA intravenously via the lateral caudal vein for 5 – 6 times at
185 10-14 days intervals in 27 days old Long Evans rats. The tail was warmed in water of 43 °C
186 prior to injection to make the vein more prominent for easy injection. The doses were
187 administered in decreasing order of 45, 40, 35, 30, 30 and 25 mg/kg. This is because the
188 older the animals the less tolerance it becomes to DMBA [18]. Moreover, the anaemia and
189 leucopenia induced by DMBA made the rats less tolerant to the chemical.

190 Leukaemia was also reported within 2.5 to 5 months in 47 % of Wistar rat fed 1-2 mg DMBA
191 twice a week. An initial dose of 200 mg/kg DMBA was administered to Long Evans rats
192 starting from 4-8 weeks of age, this was followed by set of seven doses of 10 mg/kg in 2ml
193 of sesame oil at two weeks intervals. This caused leukaemia in 72 % males, 82 % females
194 and mammary cancer in 20 % males and 66 % females. From this study it is recommended

195 that feeding of DMBA is a simple procedure for leukaemia induction, as it does not require
196 much technical skills.

197

198 **2.3 Benzo(A)pyrene (BaP) leukaemia animal model**

199 Benzo(a)pyrene is a ubiquitous polycyclic aromatic hydrocarbon that is formed from the
200 incomplete combustion at temperature ranges between 300°C to 600°C. It is mainly found in
201 residential wood burning, coal tar, tobacco smoke, automobile exhaust fumes (particularly
202 from diesel engines), and many foods including grilled meats. Its metabolites react and bind
203 to DNA causing mutations and cancer eventually. It is considered as a group 1 carcinogen
204 by the international agency for research on cancer (IARC). It is considered as the most
205 thoroughly investigated polycyclic aromatic hydrocarbons [19]. It has been reported that
206 metabolically activated benzo[a]pyrene induces cytotoxic, teratogenic, genotoxic, mutagenic,
207 and carcinogenic effects in different types of mammalian cells and tissues [20].

208 **3 CONCLUSION**

209 Different breeds of rats (including Sprague Dawley, Long Evans and Wistar rats) and mice
210 (balb c and albino) have been used as leukaemia models using various chemical
211 carcinogens including ENU, MNU, DMBA and BaP.

212 The ages of rodents at the time of chemical inoculation ranges between few days to several
213 weeks. Five days to 3 weeks in mice as well as 4-8 weeks in rats. Mortality has been
214 reported in younger mice due to acute toxicity of some of the chemicals. Male rodents are
215 most studied, perhaps due to their resistance to toxic substances more than the female.
216 However, it is recommended that female animals be also investigated to have better
217 understanding of the pathophysiology of the diseases in both sexes.

218 Various routes of administration (IP, IV and oral) have shown positive results in the animals
219 studied. In mice a total dose of 80 to 160 mg/kg body weight IP injection of ENU, given
220 singly or in divided doses at certain intervals has shown to induced leukaemia 4-7 months
221 after inoculation.

222 In rats a total of 225 mg/kg body weight IV injection of ENU in divided doses, 220 mg/kg
223 body weight IV injection of MNU in 6 divided doses and a total of 240 mg/kg body weight IP
224 injection of MNU in 4 divided doses have been shown to induce leukaemia.

225 The leukaemia induced by these chemicals in rats and mice is characterised by
226 enlargements of spleen, liver and lymph nodes, leukaemic blasts cells in both the bone
227 marrow and peripheral blood smears, cells with round or oval nuclei and occasionally with
228 indentations containing 1-3 nucleoli, among others.

229 These studies indicated that the different strains of mice and rats are susceptible to the early
230 development of chemically-induced lympho-haematopoietic system (LHS) (pre)neoplasia.
231 Consequently, these animal models appear to be suitable for use as test system in bioassay
232 protocols that adopt chemicals as initiating agents for carcinogenesis.

233

234

235

236 **REFERENCES**

237

238 1. Chang, Y.C.; Hsu, J.D.; Lin, W.L.; Lee, Y.J.; Wang, C.J. High incidence of acute
239 promyelocytic leukemia specifically induced by N -nitroso- N -methylurea (NMU) in
240 Sprague – Dawley rats. *Arch. Toxicol.* **2012**, *86*, 315–327.

241 2. DeSantis, C.; Siegel, R.; Bandi, P.; Jemal, A. Breast cancer statistics, 2011. *CA*
242 *Cancer J Clin* **2011**, *61*, 409–418.

243 3. Boffetta, P.; Kaldor, J.M. Secondary malignancies following cancer chemotherapy.
244 *Acta Oncol.* **1994**, *33*, 591–598.

- 245 4. Kohnken, R.; Porcu, P.; Mishra, A. Overview of the Use of Murine Models in
246 Leukemia and Lymphoma Research. *Front. Oncol.* **2017**, *7*.
- 247 5. Macejová, D.; Brtko, J. Chemically induced carcinogenesis: a comparison of 1-
248 methyl-1-nitrosourea, 7,12-dimethylbenzanthracene, diethylnitroso-amine and
249 azoxymethan models (minireview). *Endocr. Regul.* **2001**, *35*, 53–59.
- 250 6. Boyonoski, A.C.; Spronck, J.C.; Gallacher, L.M.; Jacobs, R.M.; Shah, G.M.; Poirier,
251 G.G.; Kirkland, J.B. Niacin deficiency decreases bone marrow poly(ADP-ribose) and
252 the latency of ethylnitrosourea-induced carcinogenesis in rats. *J. Nutr.* **2002**, *132*,
253 108–14.
- 254 7. Gerson, S.L.; Trey, J.E.; Miller, K.; Berger, N. a Comparison of O6-alkylguanine-DNA
255 alkyltransferase activity based on cellular DNA content in human, rat and mouse
256 tissues. *Carcinogenesis* **1986**, *7*, 745–749.
- 257 8. Lautier, D.; Lagueux, J.; Thibodeau, J.; Menard, L.; Poirier, G.G. Molecular and
258 biochemical features of poly (ADP-ribose) metabolism. *Mol Cell Biochem* **1993**, *122*,
259 171–193.
- 260 9. Le Rhun, Y.; Kirkland, J.B.; Shah, G.M. Cellular responses to DNA damage in the
261 absence of Poly(ADP-ribose) polymerase. *Biochem. Biophys. Res. Commun.* **1998**,
262 *245*, 1–10.
- 263 10. Zeller, W.J.; Schmähl, D. Leukemias induced by ethylnitrosourea in Wistar rats:
264 Incidence and chemotherapy. *Leuk. Res.* **1979**, *3*.
- 265 11. Law, S.; Maiti, D.; Palit, A.; Majumder, D.; Basu, K.; Chaudhuri, S.; Chaudhuri, S.
266 Facilitation of functional compartmentalization of bone marrow cells in leukemic mice
267 by biological response modifiers: An immunotherapeutic approach. *Immunol. Lett.*
268 **2001**, *76*, 145–152.
- 269 12. Bhattacharjee, B.; And, A.K.S.; Maiti, D. Role of G-CSF plus IL-15 on neutrophil
270 population in peripheral blood modulating protein tyrosine kinase activity in leukemic
271 mouse Bhaskar Bhattacharjee, Ashish Kumar Singha and Debasish Maiti. *Int. J.*
272 *Curr. Res.* **2015**, *7*, 15180–15186.
- 273 13. Singha, A.K.; Bhattacharjee, B.; Saha, B.; Maiti, D. IL-3 and GM-CSF modulate
274 functions of splenic macrophages in ENU induced leukemia. *Cytokine* **2017**, *91*, 89–
275 95.
- 276 14. da Silva Franchi, C.A., Bacchi, M.M., Padovani, C.R., & de Camargo, J.L. Thymic
277 lymphomas in Wistar rats exposed to. *Cancer Sci.* **2003**, *94*, 4–7.
- 278 15. Hutheyfa, A.H.; Hamzah, H.; S. M., R.; Sabri, J.; Mohamed Mustapha, N.; S., S.
279 Histopathological features of peripheral T-cell lymphoma in Sprague Dawley rats
280 induced with N-methyl-N-nitrosourea. *Pertanika J. Trop. Agric. Sci.* **2011**, *34*, 351–
281 361.
- 282 16. Elghetany, M.T.; MacCallum, J.M.; Davey, F.R. The use of cytochemical procedures
283 in the diagnosis and management of acute and chronic myeloid leukemia. *Clin. Lab.*
284 *Med.* **1990**, *10*.
- 285 17. Sugiyama, C.B.H. and T. Induction of leukaemia in rat by pulse doses. *Pathology*
286 **1965**, *55*, 74–81.

- 287 18. Sugiyama, T.; Osaka, M.; Koami, K.; Maeda, S.; Ueda, N. 7,12-DMBA-induced rat
288 leukemia: a review with insights into future research. *Leuk. Res.* **2002**, *26*, 1053–
289 1068.
- 290 19. Shi, Z.; Dragin, N.; Miller, M.L.; Stringer, K.F.; Johansson, E.; Chen, J.; Uno, S.;
291 Gonzalez, F.J.; Rubio, C.A.; Nebert, D.W. Oral benzo[a]pyrene-induced cancer: Two
292 distinct types in different target organs depend on the mouse Cyp1 genotype. *Int. J.*
293 *Cancer* **2010**, *127*, 2334–2350.
- 294 20. Miller, K.P.; Ramos, K.S. Impact of cellular metabolism on the biological effects of
295 benzo[a]pyrene and related hydrocarbons. *Drug Metab. Rev.* **2001**, *33*, 1–35.

297

296