Effect of *Annona ~~Muricata~~ muricata* Extract On Colon Antioxidant Status and Ketone Bodies in Blood and Urine of Cycas-Treated Rats

ABSTRACT

The effect of *Annona muricata* methanol leaf extract (AMELE) on colon antioxidant status and ~~Ketone~~ ketone bodies in blood and urine of cycas treated rats was investigated. This was to establish if there was any relationship between Soursop consumption, reduced carcinogenesis and changes in levels of ~~Ketone~~ ketone bodies. Seventy male albino rats, divided into seven groups were used for this study. Group 1 received normal diet and served as control. Group 2 received normal feed with 5% (w/w) cycas. Group 3 received ketogenic diet with 5% (w/w) cycas, Group 4 received normal feed with 100 mg AMELE/kg body wt. Group 5 received ketogenic diet with 100 mg AMELE/kg body wt. Group 6 received normal feed and 5% (w/w) cycas with 100 mg AMELE/kg body wt and Group 7 received ketogenic diet and 5% (w/w) cycas with 100 mg AMELE/kg body wt. All the treatments were administered orally for ~~twenty eight~~28 days. Using standard methods, β-hydroxybutyrate (βOH) and acetoacetate (AcAc) levels were measured in serum and urine while malondialdehyde (MDA), reduced glutathione (GSH), catalase (CAT), superoxide dismutase (SOD) and glutathione peroxidase (GPx) were assayed in the colon. Lactate dehydrogenase activities were estimated in serum, while histopathological analysis was done on colon tissues at the of 4 weeks treatment. The results showed that ~~all the groups placed on~~ ketogenic diet had significant ($p<0.05$) increase in serum and urine β-hydroxybutyrate and aceto-acetate levels compared to ~~all the~~ other groups on normal feed. *Annona muricata* extract enhanced cellular consumption and uptake of ketone bodies, whereas cycas reduced cellular consumption of ketone bodies. *Annona muricata* given either with ketogenic diet or cycas or with both ketogenic diet and cycas led to higher tissue levels of catalase, superoxide dismutase and glutathione peroxidase. Reduced glutathione levels were

UNDER PEER REVIEW

significantly ($p<0.05$) increased in ~~Group~~ group 2 (normal feed with cycas) compared to the control group. Tissue MDA levels were significantly ($p<0.05$) increased in all cycas treated groups compared to the control group with the highest levels seen in the two groups placed on normal feed and cycas (untreated/treated), Groups 2 and ~~Group~~ 6, respectively. Lactate dehydrogenase activities were ~~examined in serum and there were~~ significantly ($p<0.05$) increases in all the groups placed on ketogenic diet compared to the groups placed on normal feed. The highest activity of lactate dehydrogenase was observed in ~~Group~~ groups 3 and ~~Group~~ 5. Histo-pathological analysis revealed normal rat colon tissues in all the groups except 43% of the group 2 animals (normal feed with 5% (w/w) cycas) which showed several colonic glands with dysplastic (precancerous) changes. This invariably means that *Annona muricata* extract and/or ketogenic diet conferred protection on the colon tissues. These results therefore indicate that cycas- induced oxidative stress and tissue injury, which are early events leading to colorectal carcinogenesis, may be mitigated by the combined administration of *Annona muricata* extract and ketogenic diet.

KEYWORDS: *Annona muricata, cycas*, colon, ketogenic diets, lipid peroxidation, antioxidant, ~~Histopathology~~histopathology.

INTRODUCTION:

Incidence of cancer is ~~silently growing~~increases at exponential rates due to its nature and numerous factors associated with its development. Cancer remains one of the most dreaded diseases causing an astonishingly high death rate, second only to cardiac arrest (Shaffi *et al.*, 2009). Although overall cancer incidence rates in the developing world is half those seen in the developed world in both sexes, the overall cancer mortality rates are generally similar (Jemal *et*

UNDER PEER REVIEW

al., 2011). Cancer survival tends to be poorer in developing countries like Nigeria, most likely because of a combination of a late stage diagnosis and limited access to timely and standard treatment. The fact that conventional and newly emerging treatment ~~Assays~~ assays ~~like~~ such as chemotherapy, catalytic therapy, photodynamic therapy and radiotherapy have not succeeded in reverting the outcome of the disease to any drastic extent, has made researchers to investigate alternative treatment options (Jemal *et al*., 2011).

Over the last few decades there are accumulating evidence about the metabolic reorganisation during cancer development which has been obtained from studies on various tumor ~~types~~ (Poff *et al.,*2014). Cancer cells require unrestricted energy generation to sustain their characteristic uncontrolled proliferation (Warburg,1956). The growth of a normal mammalian cell depends on growth factor signaling and nutrient availability (Cantor and Sabatini, 2012; Dang, 2012). In certain cell models, resting cells use fatty acid oxidation as a mean~~s~~ to generate ATP, maintain membrane potentials, and turn over organelles (Wang *et al*., 2011). As the cell is induced to grow, glycolysis and glutaminolysis are induced through increased expression of glucose and amino acid transporters (Vander Heiden, 2011; Cantor and Sabatini, 2012; Dang 2012).

Most proliferating cells metabolize glucose by aerobic glycolysis rather than through the more energetically efficient oxidative phosphorylation used by normal resting cells in the presence of oxygen. It has been noted that under high conditions of high glu~~y~~cose uptake, the flux of glycolytic intermediates into branching biosynthetic pathways could be substantially increased. The dependence on glycolysis for energy production provides additional advantages to the tumor and this includes adaptation to a low oxygen environment as well as the acidification of the

UNDER PEER REVIEW

surrounding microenvironment. This promotes tumor invasion and suppresses immune surveillance (Luo *et al*., 2009).

Targeting cancer cell metabolism through glycolytic inhibition has thus emerged as a new and promising strategy to fight cancer (Seyfried, *et al,* 2003). It has been reported that ketone bodies have deleterious effects on some cancer cells (Seyfried, 2012). As a result of this, the use of ketogenic diet (KD) ~~is emerging~~ as an alternative or complementary approach to the current standard care for cancer management. Carbohydrate low ketogenic diet has been demonstrated to inhibit disease progression and promote partial remission in patients with advanced metastatic cancer from various tissue origins (Freeman *et al*., 2007; Maurer *et al*., 2011; Seyfried *et al* .,2003). On the other hand, natural products have also been targets for cancer therapy for several years. Medicinal plants are considered to be the main sources of biologically active compounds that can be used for the treatment of various ailments including cancer. One of such plants is *Annona* muricata (commonly known as soursop) which contains a novel set of phytochemicals called "annonaceous acetogenins" which have been demonstrated to exhibit remarkable cytotoxic potential against cancer cell lines (Morre *et al*.,1994; Zeng *et al*.,1996). It has an edible fruit usually called soursop due to its slightly acidic taste when ripe. *Annona muricata* is a small, upright, evergreen tree that can grow to about 4 ~~metres~~ m (13 feet) tall. Its young branches are hairy. The fruits are dark green and prickly, ovoid and can be up to 30 ~~centimetres~~ cm (12 inches) long, with a moderately firm texture. Their flesh is juicy, acidic, whitish and aromatic. Phytochemical screening of the plants showed the presence of alkaloids, carbohydrates, coumarins, flavonoids, glycosides, phenolic compounds, phytosterols, proteins, quinones, saponins, steroids and terpenoids (Vijayameena *et al*., 2013). Working on the anti-carcinogenic properties of *Annona muricata*, Maria *et al*., showed that the extracts induced

UNDER PEER REVIEW

92 necrosis of pancreatic cancer (PC) cells by inhibiting cellular metabolism (Maria *et al*.,2012).
93 Several pharmacological activities and medicinal applications of *Annona muricata* have been are
94 widely reported (Adeyemi *et al*., 2009; Adeyemi *et al*., 2010; Padmaa *et al*.,1999; Orlando *et*
95 *al*.,2010; Maria *et al*.,2012). However, their mechanisms of action has not been fully elucidated
96 beyond the fact that they inhibit the reduced form of NADH oxidase in the plasma membrane of
97 cancer cells as well as complex 1 in the election transport chain thereby starving the cancer cells
98 of ATP (Nakanishi *et al*., 2011).

99 The ~~Carcinogen~~ carcinogen used in this study was the *Cycas circinalis* plant, otherwise called
100 the Queen Sago. This plant contains alkaloids of carcinogens and also amino acids that cause
101 chronic nervous disorders. Regular consumption of the plant leads to severe health problems and
102 death. In 1941, Lorenzo and Stewart first reported the carcinogenic activity of hydrazine, the
103 hydrazine source being cycad flour. When large quantities of cycad flour were fed to rats, colon
104 adenocarcinomas ~~in the colon~~ arose in some of the animals. The carcinogen in cycad flour was
105 subsequently found to be cycasin, a form of methyl azoxymethanol (MAM) (Laqueur, 1964).
106 The carcinogen 1, 2-dimethylhydrazine (DMH) which is metabolized to MAM in the ~~liver~~
107 ~~was~~liver was found to induce cancer in inbred rats. The location of the tumor was mainly in the
108 distal portion of the colon which mimics the distribution seen in some forms of colorectal cancer
109 (Druckery and Preussmann, 1967). In 1990, the carcinogenic activity of cycad was also reported
110 when experimental animals placed on a diet composed of 5% cycad flour developed colon
111 cancer. Recently Okolie *et al*; 2013 reported the carcinogenic property of cycas after observing
112 the induction of colon carcinogenesis on experimental rats placed on a diet composed of 5%
113 cycas. Thus, this present study was aimed at examining the mechanism of action of *Annona*
114 *muricata* in relation to ketogenesis in cycas induced colon carcinogenesis in rats

Formatted: Font: Italic

UNDER PEER REVIEW

MATERIALS AND METHODS~~aterials and Methods~~

Sample Collection and Preparation~~;~~

Large quantities of fresh green leaves of *Annona muricata* were collected from trees in household gardens in Benin City, Edo State, Nigeria. Fresh leaves of *Annona muricata* were rinsed with tap water and dried at room temperature for 4 weeks. The dried leaves were ~~pulverised~~ pulverized into fine coarse powder (1.5 kg) and soaked in 2.6 L~~itres~~ of methanol for 72 hours with regular stirring. The extract was ~~collected by filtration. The residue was washed with methanol. The filtrates were combined,~~ filtered and concentrated under reduced pressure.

Fresh cycas leaflets were collected at the Ugbowo campus of the University of Benin, Benin City, Nigeria. Fresh leaves of *Cycas circinalis* were dried at room temperature and subsequently pulverised as above. The powdered leaf (2 kg) was kept in an air- tight glass jar until used.

Table 3.1: Composition of the ~~Ketogenic~~ ketogenic ~~Diet~~ diet used in this ~~Study~~study.

DIETARY COMPONENTS	AMOUNT(g/100g) (Low Carbohydrate /High Fat Diet)
Carbohydrate (Garri)	64.0~~g~~
Protein (Casein)	16.0~~g~~
Mineral and vitamin Mix	1.0~~g~~
Saturated Fat (Palm oil)	20.0~~g~~
Methionine	0.6~~g~~

The ~~Oil~~ oil used in this study was obtained from the Nigerian Institute for Oil Palm Research (NIFOR) Benin, Benin City, Nigeria. The Garri (yellow in color) was obtained from an open air market in Benin City, Nigeria.

Formatted: Font: Not Bold

Formatted: Centered, Line spacing: single

Formatted Table

Formatted: Centered, Line spacing: single

Formatted: Centered, Line spacing: single

Formatted: Centered, Line spacing: single

Formatted: Centered, Line spacing: single

Formatted: Centered, Line spacing: single

UNDER PEER REVIEW

133 **~~Animal Experiment:~~Experimental Animals**

134 A total of 70 ~~wistar~~ Wistar male rats weighing 150-to 200 g were used for this study. The rats
135 were ~~allowed two weeks for~~ acclimatization for two weeks before the commencement of ~~the~~
136 treatment. ~~They~~ The rats were maintained with tap water and fed (Growers Mash Bendel Feeds
137 and Flour Mills Ltd, Ewu, Edo State). The rats were divided into ~~7 (~~seven~~)~~ groups, each group
138 consisting of 10 rats and treated for 28 days as indicated below~~:~~.

139 Group I- Rats served as ~~Control~~control.

140 Group II- Rats + 5% (w/w) Cycas based ~~Diet~~diet.

141 Group III- Rats + 5% (w/w) Cycas based ~~Diet~~ diet + ~~Ketogenic~~ ketogenic ~~Diet~~diet

142 Group IV- Rats + AMME (100 mg/kg body wt)

143 Group V- Rats + ~~Ketogenic~~ ketogenic ~~Diet~~ diet + AMME (100 mg/kg body wt)

144 Group VI- Rats + 5% (w/w) Cycas + AMME (100 mg/kg body wt)

145 Group VII- Rats + 5% (w/w) Cycas + ~~Ketogenic~~ ketogenic ~~Diet~~ diet + AMME (100 mg/kg body
146 wt)

147

148 **Administration of *Cycas circinalis***

149 To 100 g of the rat feed, 0.5 g~~m~~ of the powdered cycas leaf was added ~~to rat feed~~ and orally
150 administered to the rats for ~~twenty-eight (~~28~~)~~ days.

151

152 **Administration of Extracts**

153 The prescribed doses of plant extracts were orally administered to the rats daily, for 28 days of
154 ~~experiment~~. Each animal was weighed. To each rat, 100 mg/kg body weight of this extract

UNDER PEER REVIEW

solution was administered. The injection dose for each animal according to their weight was determined by the following equation:

Calculated injection volume = Animals weight in kg X ~~Dose~~ dose (mg/kg)
(typically measured in ("ml") Concentration (mg/ml)

Collection of Urine

At the end of the feeding experiment, which lasted for 28 days, metabolic cages were used for urine collection.

Collection of Blood

Blood samples were collected from the anesthetized rats by cardiac puncture. The samples were collected in plain bottles.

Preparation of Serum

The blood samples collected from the rats were allowed to coagulate at room temperature and centrifuged at 3500 rpm for 15 mins at room temperature for separation of ~~Serum~~serum. The clear non-hemolysed supernatant was separated using dry pasture pipette and stored at -20°C ~~^{0} c.~~ The ~~Serum~~ serum was used for subsequent biochemical essays such as β-hydroxybutyrate, acetoacetate and lactate dehydrogenase assays.

UNDER PEER REVIEW

Biochemical Assays

Beta- hydroxybutyrate was assayed using the method described by Williamson *et al*., (1962) based on the oxidation of β-~~Hydroxybutyrate~~ hydroxybutyrate to acetoacetate by the enzyme 3-hydroxybutyrate dehydrogenase by monitoring the reduction of NAD^+ to NADH at 450 nm. The estimation of ~~Acetoacetate~~ acetoacetate (AcAc) was carried out colorimetrically at 550 nm as described by Stadtman *et al*., (1951). LDH was assayed spectrophotometrically using Randox kits, by monitoring NAD+ reduction at 340 nm as described by Klin *et al*., (1972). Catalase assay was as described by Cohen *et al*, in which the rate of decomposition of hydrogen peroxide is monitored at 480 nm. SOD was assayed by following auto-oxidation of adrenaline at 420 nm (Misra and Fridovich, 1972). Glutathione ~~Peroxidase~~ peroxidase activity was determined by measuring the production of purpurogallen from pyrogallol at 420 nm (Nyman,1959). MDA levels were measured ~~ina~~ using colorimetric reaction with thiobarbituric acid as described by Tietz, 1986. Reduced glutathione assay was done using the method as described by Tietz, 1976, in which 5,5-dithiobis (2-nitrobenzoic acid) (DTNB) with reduced glutathione (GSH) was decomposed to hydrogen peroxide and was monitored at 480nm.

Preparation of Colon Tissue Homogenates

Tissues of colon were homogenized respectively in ice cold normal saline (1:10 w/v) and centrifuged at 10,000 rpm, for 15 minutes. The supernatant was stored at 4°C~~$^{\theta}$c~~ for ~~onward~~ analysis of malondialdehyde and reduced glutathione levels, superoxide dismutase, catalase and glutathione peroxidase activities.

UNDER PEER REVIEW

Histopathological studies

The colon tissue ~~(colon)~~ were fixed with 10% formal saline solution (3-to 5 days) ~~they~~ and were later dehydrated by passing through varying (increasing) concentrations of alcohol, cleared in benzene and ~~then~~ further embedded in molten paraffin. Five micron (5 µm) cryostat sections were stained with hematoxylin and eosin dyes. The sections were examined under light microscope at high power magnifications and photomicrographs taken.

~~Results~~RESULTS AND DISCUSSION

Comment [SaA1]: Please explain about the tables in detail, rather just projecting it!

Table 1: Serum β-hydroxybutyrate and acetoacetate levels of the rats after 28 days of treatment.

	Serum β-~~Hydroxybutyrate~~ hydroxybutyrate (ng/µl)	**Serum acetoacetate (ng/µl)**
Group 1	172.66±0.70^{a}	4.27±0.60^{a}
Group 2	175.93±1.00^{ab}	4.34±0.50^{ab}
Group 3	198.98±1.00^{c}	8.63±0.60^{ch}
Group 4	170.87±0.90^{ac}	4.55±0.60^{ad}
Group 5	177.27±1.00^{ad}	7.77±1.00^{e}
Group 6	170.58±1.00^{ae}	4.18±0.70^{af}
Group 7	203.89±0.40^{e}	8.30±0.90^{gh}
One way ANOVA (p-value)	0.113	0.000

Values are represented as mean ± SEM (n=6). Means with different superscripts down the column are significantly different (p<0.05) at 95% confidence interval using paired sample students). -test.

UNDER REVIEW

Table 2: Urine β-hydroxybutyrate and acetoacetate levels of the rats after 28 days of treatment.

	Urine β-~~Hydroxybutyrate~~ hydroxybutyrate (ng/µl)	Urine acetoacetate (ng/µl)
Group 1	1.67±0.00^{a}	23.20±0.00^{a}
Group 2	2.70±0.00^{b}	30.00±0.00^{b}
Group 3	4.00±0.00^{c}	70.70±0.00^{c}
Group 4	1.60±0.00^{a}	33.70±0.00^{d}
Group 5	5.70±0.00^{d}	83.50±0.00^{e}
Group 6	0.90±0.00^{e}	26.50±0.00^{f}
Group 7	2.70±0.00^{f}	84.20±0.00^{g}
One way ANOVA (p-value)	0.000	0.000

Values are represented as mean±SEM (n=6). Means with different superscripts down the column are significantly different (p<0.05) at 95% confidence interval using paired sample students' t-test..

Table 3: Colorectal tissue antioxidant enzyme activities and reduced glutathione levels of rats after 28days of treatment.

	Tissue reduced glutathione (mmole/mg)	Tissue Superoxide dismutase $(U/mg)10^{-3}$	Tissue Catalase (U/mg)	Tissue glutathione peroxidase (U/mg)
Group 1	0.33±0.01^{ac}	0.21±0.03^{ac}	95.29±2.00^{ac}	0.33±0.01^{abg}
Group 2	0.49±0.10^{bh}	0.12±0.03^{b}	91.43±0.80^{b}	0.3±0.06^{b}
Group 3	0.40±0.07^{c}	0.30±0.07^{ch}	105.42±0.40^{ch}	0.43±0.07^{ch}
Group 4	0.37±0.08^{d}	0.09±0.03^{d}	86.14±0.70^{d}	0.27±0.04^{d}
Group 5	0.51±0.10^{e}	0.32±0.10^{e}	142.14±0.10^{e}	0.42±0.07^{eh}
Group 6	0.69±0.10^{fhi}	0.12±0.04^{fhi}	162.14±0.40^{f}	0.51±0.09^{f}
Group7	0.56±0.10^{gi}	0.21±0.06^{gi}	84.57±2.00^{gh}	0.28±0.05^{bg}
One-way ANOVA (p-value)	0.492	0.058	0.089	0.099

Values are represented as mean ± SEM (n=6). Means with different superscripts are significantly different (p<0.05) at 95% confidence interval.

UNDER PEER REVIEW

Table 4: Colorectal tissue level of lipid peroxidation marker (MDA) and serum lactate dehydrogenase activity of rats after 28days of treatment.

	MDA (x 10^{-3} nmoles/ g of fresh wt).	Serum lactate dehydrogenase (IU/ml)
Group 1	1.24±0.71[a]	583.89±0.90[a]
Group 2	2.25±0.20[b]	478.70±0.40[b]
Group 3	1.36±0.20[ch]	1758.28±2.00[ch]
Group 4	1.21±0.20[d]	518.17±0.20[di]
Group 5	1.72±0.05[e]	1780.24±0.80[eh]
Group 6	1.97±0.40[f]	519.99±0.50[fi]
Group 7	1.29±0.10[gh]	741.61±0.80[g]
One-way ANOVA (p-value)	0.000	0.000

Values are represented as mean±SEM (n=6). Means with different superscripts are significantly different ($p<0.05$) at 95% confidence interval down the column by paired sample students' t-test.

~~**HISTOPATHOLOGICAL RESULTS**~~

The histopathological results obtained from this study showed normal mucosal and muscular layers in all the groups placed on ~~Ketogenic~~ ketogenic diet, treated or untreated with cycas and or *Annona muricata* except the group placed on Normal diet treated with cycas only.

This can be seen in the ~~Plates~~ Figures are arranged as below.~~:~~

Comment [SaA2]: Need explanation for every images presented! What are the changes before and after!

UNDER PEER REVIEW

238

239

240

241

242

243

244

245

246

247

248

249

250

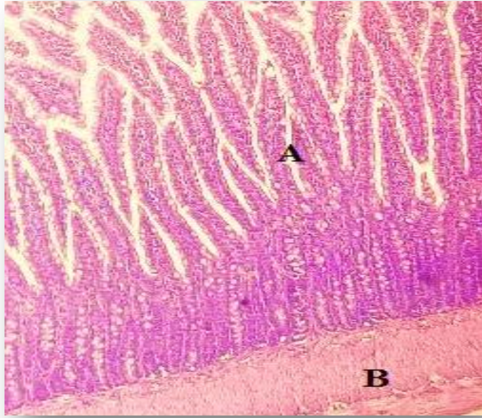
251

252

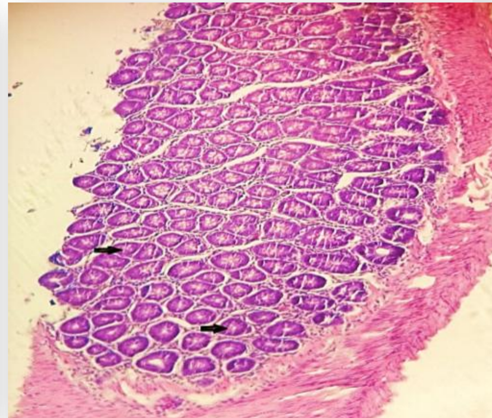
253

254

255



~~Plate~~ Figure 1: Longitudinal section of rat intestine from control group. Intact mucosal (**A**) and muscular (**B**) layers are evident.The mucosa is seen here as finger-like projections of its lamina propria covered by normal columnar epithelial cells and mucus-secreting goblet cells. The muscularispropria is the muscle layer very well delineated at the bottom of the micrograph, propping up the mucosa. (H and E; X 100).



~~Plate~~ Figure 2: Transverse section of the colon from a control rat. Normal intestinal glands are evident in the mucosa, each being shown here as circular (tubular) structures (arrows) lined by mucus-secreting columnar epithelial cells. The underlying smooth muscle layer (muscularispropria) is also ~~shown (~~shown (H and E; X 100).

Formatted: Indent: Left: 0", First line: 0"

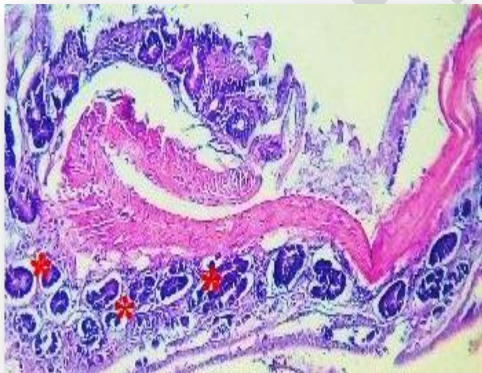


Plate 3: Section of the colon from a rat treated with Cycas only. (H & E; x 100). Intestinal glandular epithelial dysplasia (pre-cancerous change) is evident across the glands of the mucosa (red asterisks), the epithelial lining cells of which obviously exhibit nuclear enlargement, hyperchromasia and loss of mucin secretion. These pre-cancerous changes were seen in only about 43% of the rats ~~belonging to this group.~~and ~~The~~ the underlying smooth muscular layer is also shown.

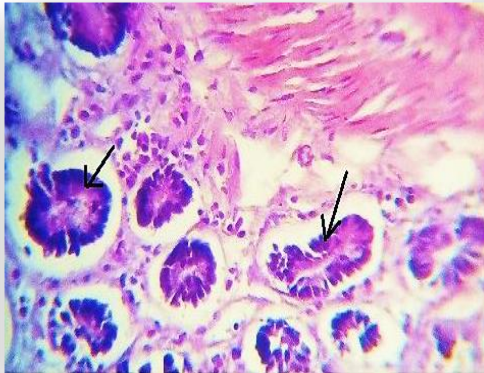
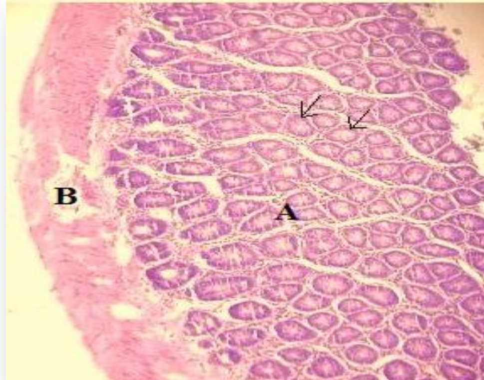
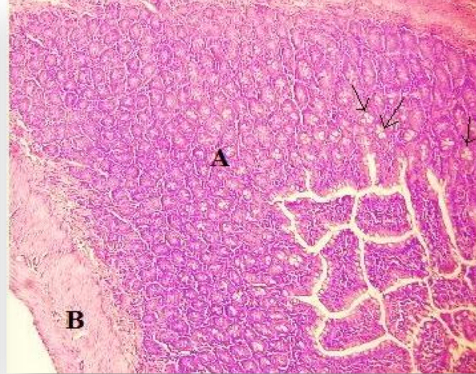


Plate 4: Section of the colonic mucosa from a Cycas-only treated rat (Hand E; x 100). Glands (arrows) lined by dysplastic epithelial cells with nuclear disarray, enlargement and hyperchromasia are evident. There is loss of mucin secretion and absence of goblet cells. These pre-cancerous changes were seen in only about 43% of the rats belonging to this group.

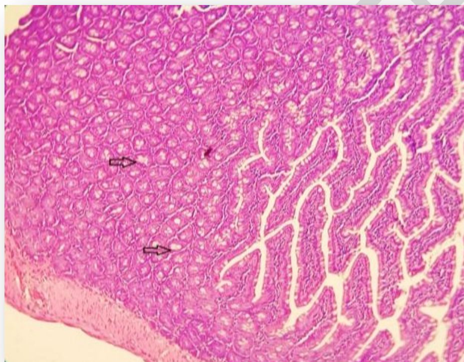
PEER REVIEW



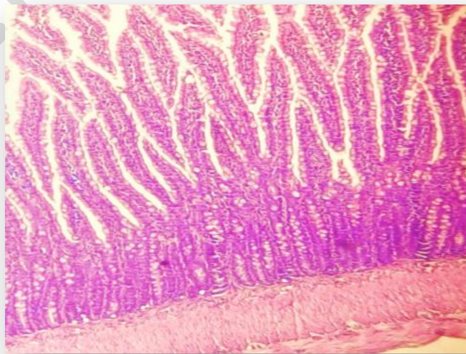
~~Plate~~ **Figure 5:** Section of the colon from a cycas-only treated rat (H&E; x100).The mucosa (A) contains numerous normal, transversely sectioned tubular glands (arrows) within its lamina propria. Each gland has a central lumen surrounded by columnar epithelial lining cells. The muscular (B) layer is also well shown. Such normal histology was seen in 57% of this group.



~~Plate~~ **Figure 6:** Section of the colon from a cycas and ketogenic diet treated rat (H&E; x100). A normal mucosa (A) with normal glands lined by columnar and goblet cells (arrows), a normal muscularis layer (B) and an inconspicuous layer of submucosa inbetweenthe mucosa and muscularis~~,~~ are evident.

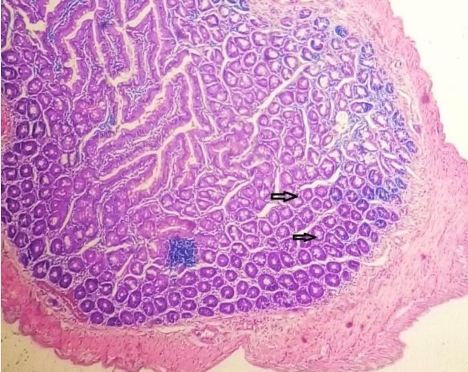


~~Plate~~ **Figure 7:** Section taken from an extract only treated rat (H& E; x100).Numerous normal mucous glands (arrows) lined by normal columnar epithelial and goblet cells are evident in this view. The right of the field shows the mucosa disposed as finger-like structures of lamina propria covered by columnar and goblet cells. A section of the muscular layer is seen at the bottom left.

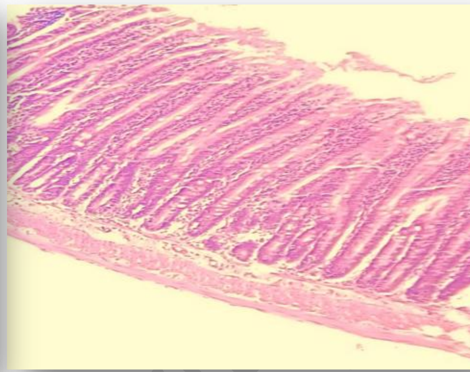


~~Plate~~ **Figure 8:** Section taken from a rat placed on ketogenic diet and extract (H&E: x100).Normal histology is evident. The mucosa above is displayed in numerous fingerlike projections of the lamina propria covered by columnar lining cells and mucin-secreting goblet cells. The muscularis propria props up the mucosa as shown at the bottom of the micrograph.

UNDER PEER REVIEW



~~Plate~~ Figure 9: Section of colon from a cycas and extract treated rat (H&E; x100). The normal mucosa with normal glandular architecture (arrows) is shown here surrounded by the normal muscular layer.



~~Plate~~ Figure 10: Section of colon from a cycas, extract and ketogenic diet treated rat (H&E; x100). Numerous straight, tubular glands are seen in this normal mucosa, and they are lined by normal columnar and goblet cells. A thin layer of submucosa separates the mucosa above from the muscularis propria beneath.

~~Discussion~~

Cancer chemoprevention ~~by~~ using antioxidant approaches has been ~~suggested to offer~~ a good potential in providing important fundamental benefits to public health and is now considered by many clinicians and researchers as a key strategy for inhibiting, delaying or even reversal of process of carcinogenesis (Ahmeida *et al.,* 2005). Moreover, knowledge and application of such potential activities in reducing oxidative stress *in vivo* has prompted the search for potent and cost effective antioxidants from various plant sources. It has been strongly suggested that colorectal carcinogenesis is associated with intense oxidative stress and the progression of the cancer is certain in the face of increased imbalance in the oxidative-antioxidative disorder (Skrzydlewska *et al*., 2001).

UNDER PEER REVIEW

294 The present study assays the tissue-protective effect of *Annona muricata* extract against
295 histopathological damage in cycas-–induced oxidative-stressed rats. Results from ~~table~~ Table 1
296 shows the ~~Serum~~ serum levels of β-hydroxybutyrate and acetoacetate levels of the rats after 28
297 days of treatment. Results ~~from the show~~showed significant increase ($p<0.05$) in serum ~~Ketone~~
298 ketone body (β -hydroxybutyrate levels and –acetoacetate) profiles of the groups placed on
299 ~~Ketogenic~~ ketogenic diet (groups, III, V and VII) compared to the groups placed on normal diet
300 (Table 1). ove, it could be noticed that rats fed with ketogenic diets (groups, III, V and VII)
301 showed higher ketone bodies (β -hydroxybutyrate levels and acetoacetate). This is true because
302 Ketogenic diet leads to the production of ketone bodies, such as β-hydroxybutyrate and
303 acetoacetate, which can be used as an alternative energy source. Within the groups cycas treated
304 groups, group VI has the lowest ketone body profile.

305

306 Results from ~~table~~ Table 2 show a significant increase ($p<0.05$) in urinary ~~Ketone~~ ketone body
307 profiles in the groups (III, V and V11) placed on ~~Ketogenic~~ ketogenic diet compared to the
308 groups (I, II, IV and VI) placed on normal diet as shown in Table 2 ~~below~~. Within the cycas
309 treated groups (II, VI and VII), the untreated carcinogenic group (I) presented the lowest urinary
310 ketone body. This reduced presence of urinary ketone body as a result of inability of the
311 cancerous cells to efficiently process ketone bodies for energy. Ketones also slow the
312 proliferation of tumo~~u~~r cells. This reduced ketone bodies in the urine is also evident in the
313 reduced serum ketone body profile within the group as shown in ~~table1~~Table 1. Although ketone
314 bodies are efficient energy substrates for healthy extra_hepatic tissues, cancer cells cannot
315 effectively use them for energy (Veech *et al.,* 2001). This increase in the ketone body of the rats
316 treated with ketogenic diet corroborates with the results of a “classical” ketogenic diet described

Formatted: Don't adjust space between Latin and Asian text, Don't adjust space between Asian text and numbers

Comment [SaA3]: What’s this?

Formatted: Highlight

Formatted: Highlight

UNDER PEER REVIEW

by Russell Wilder at the Mayo clinic (Wilder, 1921). Studies by ~~Poff in 2104 (~~Poff *et al.,* 2014~~)~~ shows that ~~Ketone~~ ketone supplementation elevated blood ketones which is similar with the present study. Hence, from the present study, we can hypothesize that dietary administration of ketone body precursors would inhibit disease progression *in vivo*.

Among the measures that are utilized ~~-~~by cells to counteract the deleterious effects of lipid peroxidation is ~~the~~ alteration of the activity of antioxidant enzymes such as catalase, peroxidases and superoxide dismutase (Bhor *et al*., 2004). Catalase is an important cellular antioxidant enzyme that defends against oxidative stress found in the peroxisomes of most aerobic cells. It serves to protect the cell from toxic effects of high concentrations of hydrogen peroxide by ~~catalysing~~catalyzing its decomposition into molecular oxygen and water (Shangari 2006). Thus, estimation of catalase activity is a good indicator of oxidative stress in experimental animals. Superoxide dismutase is a metalloenzyme that scavenges superoxide anions and exists as isoforms classified on the basis of their relative requirement for manganese, copper and zinc to maintain structural and functional integrity (Sasaki *et al*., 2000). The results as seen in Table 3 ~~above,~~ shows a significant increase in colon tissue ~~Superoxide~~ superoxide dismutase, ~~Glutathione~~ glutathione peroxidise and catalase activities in all the ~~Ketogenic~~ ketogenic diet/ *Annona muricata* treated groups compared to the ~~Normal~~ normal diet/ *Annona muricata* treated groups.

The Anti~~-~~oxidant enzyme system is one of the inherent protective mechanisms in living organisms. *Annona muricata* has been reported to enhance anti~~-~~oxidative enzyme system~~s~~ in experimental animals. The observed synergistic relationship between *Annona muricata* and

UNDER PEER REVIEW

ketogenic diet in this study was also confirmed in the levels of anti~~-~~oxidant enzyme activity exhibited between group_4 (normal diet/extract treated) and group_5 (ketogenic diet/extract treated). In this study, there was a significant (P < 0.05) increase in antioxidant enzyme activity in group_5 compared to group_4. Some studies, however, reported that ketogenic diet~~, (~~with high fat, low carbohydrate diets~~)~~ can modulate the response of some antioxidant enzymes (Jodynis-Liebert and Murias, 2002; Dhuley *et al.,* 1999; Ruiz-Gutierrez *et al.,* 1999).

The animals in group 2 (normal diet, cycas treated), however showed lower levels of antioxidant activity compared to control rats but they showed higher levels of reduced glutathione compared to control rats. This is not uncommon as oxidative stress can cause the up-regulation of antioxidant enzymes and increase in reduced glutathione levels that render cells more resistant to subsequent oxidative insult (Halliwell, 2000). Over-expression of glutathione has been reported in both animal and human tumors (Yang *et al*., 1997; Skrzydlewska *et al.,* 2001; Balasenthil., 2000).

Results of the antioxidants levels of the rats in the present study shows (~~table~~ Table 3) that ~~they rats treated with~~ only the *Annona muricata* extract (group IV) presented the lowest antioxidant levels. This was similar to the levels shown by the normal rats (group I), indicating that *Annona muricata* did not in any way cause oxidative damage. Ketogenic diet has been reported to stimulate the cellular endogenous antioxidant system with the activation of nuclear factor erythroid-derived 2 (NF-E2)-related factor 2 (Nrf2), the major inducer of detoxification genes

UNDER PEER REVIEW

(Pinto, *et al.,* 2018). This could probably be the same reason why the groups treated with ketogenic diets presented higher antioxidant levels.

Results seen in Table 4 above show a significant increase in the malondialdehyde content in the cycas-treated groups placed on ~~Ketogenic~~ ketogenic diet. The values of ~~Lactate~~ lactate dehydrogenase activity was significantly higher in groups placed on ~~Ketogenic~~ ketogenic diet (treated/untreated) compared to the groups placed on normal diet (treated/untreated).

Members of the *Cycas* family have been shown to contain cycasin which has the ability to cause the formation of agents like azoxymethane which have free radical activity (Rosenberg *et al*., 2009). This may account for the elevated amount of malondialdehyde-an index of lipid peroxidation in the experimental animals exposed to cycads only compared with the controls. Lipid peroxidation is a well-established mechanism of cellular injury and therefore used as an indicator of oxidative stress which is characterised by an increase in malondialdehyde concentration (MDA). In this study, the concentration of MDA was markedly increased in the cycas-treated group which is significantly different (p<0.05) from the control group and the other groups. This is similar to the findings of Okolie *et al*., (2013); Lolodi and Eriyamremu, (2013); Eriyamremu *et al*., (2007) in which an increase in MDA concentration after cycas administration was reported. The result of the ~~Lipid~~ lipid ~~Peroxidation~~ peroxidation assay therefore shows the oxidative stress effect caused by cycas administration. Within the cycas groups (II, III, VI and VIII) treated, group VII presented the lowest MDA level thereby suggesting a possible synergistic effect of both extracts.

UNDER PEER REVIEW

384 Histological results ~~from this study~~ showed that~~,~~ colon sections of cycas only treated rats (Group
385 II) showed evidence of pre-cancerous changes when compared to the untreated controls shown in
386 ~~plate~~Figures 3 and 4. Section of the colon from a rat treated with ~~Cycas~~ cycas only showed that
387 there was evident intestinal glandular epithelial dysplasia (pre-cancerous change) across the
388 glands of the mucosa. The epithelial lining cells of which obviously exhibit nuclear enlargement,
389 hyperchromasia and loss of mucin secretion. Sections from rats from other groups presented
390 normal morphological appearance. As shown in the ~~plates above~~Figures, as treatment was
391 introduced; the tissue (Groups VI, and VII) sections were evidently restored to normal. It seems
392 *Annona muricata* has tissue-protective effect which can be observed by its ability to restore and
393 reverse the damaged tissues. Similar effects were reported by earlier studies on *Annona*
394 *muricata.* Working on the evaluation of ~~Anticancer~~ anticancer ~~Activity~~ activity of ~~Annona~~
395 annona muricatain 1, 2-~~Dimethyl~~ dimethyl ~~Hydrazine~~ hydrazine ~~Induced~~ induced ~~Colon~~ colon
396 ~~Cancer~~cancer, Venkateshwarlu *et al,* (2014) reported that the ethanol extract of *Annona muricata*
397 it has shows potent anti–cancer activity. This was also similar to reports presented by
398 Moghadamtousi *et al*. in 2014. Using HCT-116 and HT-29 cells and MTT and LDH assays, it
399 has been shown that cells treated by ethanol extracts of *A.* ~~*muricata* arrested~~*muricata* arrested in
400 G1 cell cycle phase and induced to apoptosis cascade. Ethanol extracts of *A. muricata* -treatment
401 caused excessive accumulation of ROS followed by disruption of MMP, cytochrome c leakage
402 and activation of the initiator and executioner c as passes in both colon cancer cells
403 (Moghadamtousi *et al.,* 2014a). It's also reported that ethyl acetate extract of *A. muricata*
404 inhibited the proliferation of A549 cells, leading to cell cycle arrest and programmed cell death
405 through activation of the mitochondrial-mediated signaling pathway with the involvement of the
406 NF-κB signalling pathway (Moghadamtousi *et al.,* 2014b).

Formatted: Font: Italic

Formatted: Font: Italic

UNDER PEER REVIEW

The anti-proliferative activity of *A. muricata* extract had been also reported elsewhere. A comprehensive study showed that potentially strong antiproliferation and apoptosis was achieved through disruption of MMP (Membrane mitochondrial potential), reactive oxygen species (ROS) generation and G0/G1 phase cell mitigate Human promyelocytic leukemia (HL-60 cells) cell culture (Pieme *et al.,* 2014). Reports also showed that treatment with *Annona muricata* improved the histological changes of breast tissue and reduced the proliferative indexes of their cells (Sulistyoningrum *et al.* 2017). This was in line with previous *in vitro* studies; the leaves of soursop had *in vitro* anticancer activity on T47D cell lines (Rachmani *et al.,* 2012), human adenocarcinoma cell MCF-7 (Ko *et al.,* 2011), human carcinoma cell (MDA-MB-435S) or human immortal keratino cytes (HaCaT) (George *et al.,* 2012).

Formatted: Font: Italic

Formatted: Font: Italic

Formatted: Font: Italic

The chemo preventive effects of *Annona muricata* might be attributed to the presence of a class of compound obtained from Annona species, acetogenins. It has been reported that the main anti-tumorous compound, annonacin was effective against *in vitro* cancer cell lines as well as *in vivo* lung cancer (Venkateshwarlu *et al* 2014).

CONCLUSIONS

This study reveals that *Annona muricata* increased cellular uptake consumption of ketone bodies while *Cycas circinalis* appeared to have the opposite effect. This property exhibited by *Annona muricata could* be exploited as a therapeutic tool in the management and treatment of cancer. The anti-oxidative properties of *Annona muricata* were evident in this study. ~~From the histology results, ketogenic~~ Ketogenic diet and *Annona muricata* protected the cells from structural damage~~. Together,~~and they exerted even more protection on these experimental animals.

UNDER PEER REVIEW

REFERENCES

Adeyemi, D.O., Komolafe, O.A., Adewole, O.S., Obuotor, E.M. and Ademowo, T.H. (2009). Anti-~~Hyperglycemic~~ hyperglycemic ~~Activities~~ activities of *Annona muricata* (Linn). *Afr. J. Trad. CAM.* 6(1): 62-69.

Adeyemi, D.O., Komolafe, O.A., Adewole, O.S., Obuotor, E.M., Abiodun, A.A. and Adenowo, T.K. (2010). Histomorphologocal and ~~Morphometric~~ morphometric ~~Studies~~ studies of the ~~Pancreatic~~ pancreatic ~~Islet~~ islet ~~Cells~~ cells of ~~Diabetic~~ diabetic ~~Rats~~ rats treated with extracts of *Annona muricata. Folia. Morphol. (Warsz).* 69(2): 92-100.

Almeida, V. L., Leitão, A., Reina, L. C. B., Montanari, C. A., Donnici, C. L., and Lopes, M. T. P. (2005). Cancer and cell cicle-specific and cell cicle nonspecific anticancer DNA-interactive agents: an introduction; *Química Nova*, *28*(1), 118-129. https://dx.doi.org/10.1590/S0100-40422005000100021

Balasenthil, S., Saroja, M., Ramachandran, C. R. and Nagini, S. (2000). Of Humans and ~~Hamsters~~hamsters: Comparative ~~Analysis~~ analysis of ~~Lipid~~ lipid ~~Peroxidation~~ peroxidation ~~Glutathione~~ glutathione and ~~Glutathione~~glutathione-~~Dependent~~ dependent ~~Enzymes~~ enzymes during ~~Oral~~ oral ~~Carcinogenesis~~carcinogenesis. *Er. J. Oral. Maxillofac.Surg*.38:267-70.

Bhor, V.M., Raghuram, N and S. Sivakami, S. (2004). Oxidative damage and altered antioxidant enzyme activities in the small intestine of streptozotocin-induced diabetic rats. Int. J. Biochem. Cell. Biol., 36: 89-97.

Cantor, J.R. and Sabatini, D.M. (2012). Cancer cell metabolism: One hallmark, many faces. *Cancer. Discov.* 2:881-898.

Formatted: Font: Italic

UNDER PEER REVIEW

Dang, C.V. (2012). Links between metabolism and cancer. *Genes. Dev.* 26:877-890.

Dhuley, J. N. (1999). Anti-~~Oxidant~~ oxidant ~~Effects~~ effects of ~~Cinnamon~~ cinnamon *Cinnamommumverum* ~~Bark~~ bark and ~~Greater~~ greater *Cardamon Amommum Subulatum* seeds in ~~Rats~~ rats fed ~~High~~ high ~~Fat~~ fat ~~Diet~~diet. *Ind. J .Exp. Biol*. 37:238-42.

Druckrey, H., Preussmann, R. and Matzkies, F. (1976).Selective ~~Production~~ production of ~~Intestinal~~ intestinal ~~Cancer~~ cancer in ~~Rats~~ rats by 1, 2 – Dimethylhydrazine.*Natur wissens chaften*54:285-286.

Eriyamremu, G.E., Asagba, S.O. Uanseoje, S.O. Omeregie, S.E. and. Omofoma, C.O (2007). Colonic lipid peroxidation, nuclear membrane ATPase and stress enzymes in rats fed a Nigerian-~~Like~~ like ~~Diet~~ diet and ~~Cycas~~cycas. J. Boil. Sci., 7: 526-531.

Freeman, J. M., Kossoff, E. H. and Hartman, A. L. (2007). The Ketogenic ~~Diet~~diet: One ~~Decade~~ decade ~~Later~~later. *Pediatrics,* 119(3):535-43.

George, V.C., Kumar, D.R.N., Rajkumar, V., Suresh, P.K., Kumar, R. A. (2012) Quantitative ~~Assessment~~ assessment of the ~~Relative~~ relative ~~Antineoplastic~~ antineoplastic ~~Potential~~ potential of the n-butanolic ~~LeafExtract~~ leaf extract of *Annona ~~Muricata~~ muricata* Linn. in ~~Normal~~ normal and ~~Immortalized~~ immortalized ~~Human~~ human ~~Cell~~ cell ~~Lines~~lines. *Asian Pacific J Cancer Prev*, ;13 (2): 699-704.

Halliwell, B. (2000). The ~~Antioxidant~~ antioxidant ~~Paradox~~paradox.*Lancet.*355:1179-80.

Jermal, A., Bray, F., Center, M.M., Ferlay, J., Ward, E. and Forman, D. (2011). “Global ~~Cancer~~ cancer ~~Statistics~~statistics”. *Cancer Journal for Clinicians.*61(2): 134.

UNDER PEER REVIEW

Jodynis-Liebert, J. and , Murias, M.() Modulation of antioxidant defence system by dietary fat in rats intoxicated with o-toluidine. *Hum Exp Toxicol.* 21(12):659-65.

Klin, Z.(1972). Kinetic determination of ~~Lactate~~ lactate dehydrogenase as ~~Recommended~~ recommended by the German Clinical Chemistry Society (DGKC) Biochemistry.*Chem.Klin.Biochem*.I.8:658.

Ko, Y.M., Wu, T.Y., Wu, Y.C., Chang, F.R., Guh, J.Y., Chuang, L.Y. (2011). Annonacin induces cell cycle-dependent growth arrest and apoptosis in estrogen receptor-α-related pathways in MCF-7 cells. *J Ethnopharmacol*, ; 137 (3):1283-1290.

Laqueur, G. (1964). Carcinogenic ~~Effect~~ effect of ~~Cycad~~ cycad ~~Meal~~ meal and ~~Cycasin~~cycasin, ~~Methylazoxymethanol~~ methylazoxy methanol glycoside in ~~Rats~~ rats and ~~Effects~~ effects of ~~Cycasin~~ cycasin in ~~Germ~~ germ – ~~Free~~ free ~~Rats~~rats. *Fed. Proc*.23:1386-1387.

Lolodi O. and Eriyamremu, G.E. (2013). Effect of ~~Methanolic~~ methanolic ~~Extract~~ extract of *Vernonia amygdalina* (~~Common~~ common ~~Bitter~~ bitter ~~Leaf~~leaf) on ~~Lipid~~ lipid ~~Peroxidation~~ peroxidation and ~~Antioxidant~~ antioxidant ~~Enzymes~~ enzymes in ~~Rats~~ rats ~~Exposed~~ exposed to ~~Cycasin~~cycasin. *Pakistan Journal of Biological Sciences, 16: 642-646.*

Luo, J., Solimini, N. L. and Elledge, S.J. (2009). Principles of cancer therapy: Oncogene and ~~Non~~non-~~Oncogene~~ oncogene addiction. *Cell*.136:823-837.

Maria, P.T., Satyanarayana, R., Vinee, P., Poomy, P., Suhasini, J., Erik, D. M., Sonny, J., Pankaj, S., Apar, K.G. and Surinder. K.B. (2012). Graviola-~~Novel~~ novel ~~Promising~~ promising ~~Natural~~ natural derived drug that inhibits tumorigenicity and metastasis of ~~Pancreatic~~

UNDER PEER REVIEW

493 pancreatic ~~Cancer~~ cancer ~~Cell~~ cell *in vitro* and *in vivo* through ~~Alternating~~ alternating
494 cell metabolism. *Cancer Cell Letters.* 323: 29-40.
495 Maurer, G. D. (2011). Differential ~~Utilization~~ utilization of ~~Ketone~~ ketone ~~Bodies~~ bodies by
496 ~~Neurons~~ neurons and ~~Glioma~~ glioma cell Lines: A ~~Rationale~~ rationale for ~~Ketogenic~~
497 ketogenic ~~Diet~~ diet as ~~Experimental~~ experimental ~~Glioma~~ glioma ~~Therapy~~therapy.
498 *BMC Cancer*.11:315.
499 Misra, H.P. and Fridovich, I.(1972). The ~~Role~~ role of ~~Superoxide~~ superoxide ion in the ~~Anti~~anti-
500 oxidation of ~~Epinephrine~~ epinephrine and a ~~Simple~~ simple ~~Assay~~ assay for ~~Superoxide~~
501 superoxide ~~Dismutase~~dismutase.*J.Biol.Chem.*247:3170-3175.
502 Moghadamtousi, S. Z., Kadir, H. A., Paydar, M., Rouhollahi, E. and Karimian, H., (2014a)
503 *Annona muricata* leaves induced ~~Apoptosis~~ apoptosis in A549 cells through
504 ~~Mitochondrial~~mitochondrial-~~Mediated~~ mediated ~~Pathway~~ pathway and ~~Involvement~~
505 involvement of NF-XB. *BMC Complementary and Alternative Medicine*.14:299.
506 Moghadamtousi, S.Z., Kadir, H.A., Paydar, M., Rouhollahi, E., Karimian, H. (2014b) Annona
507 muricataleaves induced apoptosis in A549 cells through mitochondrial-mediated
508 pathway and involvement of NF-κB. *BMC Complement Altern Med*, ; 14 (1): 1-13.
509 Morré, D.J., de Cabo, R., Farley, C., Oberlies, N.H., McLaughlin, J.L.(1994) Mode of action of
510 bullatacin, a potent antitumor acetogenin: inhibition of NADH oxidase activity of HeLa
511 and HL-60, but not liver, plasma membranes. Life Sci. 1994;56(5):343–348.
512 Nishisho, T., Hata, K., Nakanishi, M., Morita, Y., Sun-Wada, G., Wada, Y., Yasui, N., and
513 Yoneda, T. (2011). The *a*3 ~~Isoform~~ isoform ~~Vacuolar~~ vacuolar ~~Type~~ type H^+-ATPase
514 ~~Promotes~~ promotes ~~Distant~~ distant ~~Metastasis~~ metastasis in the ~~Mouse~~ mouse B16

Formatted: Font: Italic

UNDER PEER REVIEW

515 ~~Melanoma~~ melanoma ~~Cells~~cells: *Mol Cancer Res* 9)(7) 845-855; DOI:
516 10.1158/15417786.MCR-10-0449

517 Nyman,M.(1959). Serum ~~Haptoglobin~~haptoglobin: Methodological and ~~Clinical~~ clinical Studies.
518 *Scand .J .Clin.Lab Invest*.11:39-39.

519 Okolie, N.O., Agu, K., Eze, G.I. (2013)~~-~~. Protective ~~Effect~~ effect of ~~Ethanolic~~ ethanolic ~~Leaf~~ leaf
520 extract of *Annona muricata* Linn on some early events in ~~Cycas~~cycas-induced
521 colorectal carcinogenesis in rats. *JSPI*.2(4)14-21.

522 Orlando, V., Glauciemar, D.V., Jose de Jesus, R.G.D., Celia, H.Y. and Maria, S.A. (2010).
523 Antinociceptic and ~~Anti~~anti-inflammatory ~~Activities~~ activities of the ~~Ethanol~~ ethanol
524 ~~Extract~~ extract of *Annona muricata*. L. ~~Leaves~~ leaves in ~~Animal~~ animal ~~Models~~models.
525 *Int. J. Mol. Sci.* 11: 2067-2078.

526 Padmaa, B.P., Chansouria, J.P.N. and Khosaa, R.L. (1999). Hepatoprotective ~~Activity~~ activity of
527 *Annona muricata* Linn and *Polyalthia ~~Cerasoide~~ cerasoide Ancient Science of Life*.
528 19:7-10.

529 Pieme, C. A., Santosh, G. K., Tekwu, E. M., Askun, T., Aydeniz, H., Ngogang, J. Y., and
530 Saxena, A. K. (2014). Fruits and barks extracts of *Zanthozyllum heitzii* a spice from
531 Cameroon induce mitochondrial dependent apoptosis and Go/G1 phase arrest in human
532 leukemia HL-60 cells. *Biological research*, *47*(1), 54. doi:10.1186/0717-6287-47-54

533 Pinto, A., Bonucci, A., Maggi, E., Corsi, M., and Businaro, R. (2018). Anti-~~Oxidant~~ oxidant and
534 ~~Anti~~anti-Inflammatory ~~Activity~~ activity of ~~Ketogenic~~ ketogenic ~~Diet~~diet: New
535 ~~Perspectives~~ perspectives for ~~Neuroprotection~~ neuroprotection in Alzheimer's
536 ~~Disease~~disease. *Antioxidants (Basel, Switzerland)*, *7*(5), 63. doi:10.3390/antiox7050063

Formatted: Font: Italic

Formatted: Font: Italic

Formatted: Font: Italic

Formatted: Font: Italic

UNDER PEER REVIEW

537 Poff, A. M., Ari, C., Arnold, P., Seyfried, T. N. and D'Agostino, D. P. (2014). Ketone ~~SupplementationDecreases~~ supplementation decreases ~~Tumor~~ tumor ~~Cell~~ cell ~~Viability~~ viability and ~~Prolongs~~ prolongs ~~Survival~~ survival of mice with ~~Metastatic~~ metastatic ~~Cancer~~cancer. *Int. J. Cancer.*135:1711-1720.

541 ~~Poff, A.M. Ari, C. Arnold,P., Seyfried, T.N. and D 'Agostino, D.P. (2014) Ketone supplementation decreases tumor cell viability and prolongs survival of mice with metastatic cancer. *Int. J. Cancer:* 135, 1711–1720~~

544 Rosenberg, D.W., Giardina, C. and Tanaka T. (2009). Mouse models for study of colon carcinogenesis. Carcinogenesis, 30: 183-196.

546 Ruiz-Gutiérrez, V., Pérez-Espinosa, A., Vásquez, C. M. and Santamaria, C. (1999). Effects of ~~Dietary~~ dietary ~~Fat~~ fat (fish, olive and high-oleic-acid sunflower oils) on ~~Lipid~~ lipid ~~Composition~~ composition and ~~Antioxidant~~ antioxidant ~~Enzymes~~ enzymes in ~~Rat~~ rat ~~Liver~~liver. *Br. J. Nutr.* 82:233-41.

550 Sasaki, H., Akamatsu, H. and . Horio, T. (2000). Protective role of copper, zinc superoxide dismutase against UVB-induced injury of the human keratinocyte cell line HaCaT. *J. Invest. Dermatol.*, 114: 502-507.

553 Seyfried, T. N. (2012). Metabolic ~~Management~~ management of cancer as a ~~Metabolic~~ metabolic ~~Disease~~disease; On the ~~Origin~~origin, ~~Management~~ management and ~~Prevention~~ prevention of cancer. John Wiley and Sons, Hoboken, NJ, pp. 291-354.

556 Seyfried, T. N., Sanderson, T. M., El-Abbadi, M. M., McGowan, R. and Mukherjee, P. (2003).Role of Glucose and ~~Ketone~~ ketone ~~Bodies~~ bodies in the ~~Metabolic~~ metabolic

Formatted: Strikethrough

UNDER PEER REVIEW

~~Control~~ control of ~~Experimental~~ experimental ~~Brain~~ brain ~~Cancer~~cancer. *Br. J. cancer.* 89:1375-1382.

Shafi, G., Anjana, M., Hasan, T., Alshatwi, A., Tyothy, A., and Lei, D.K. (2009). Induction of ~~Apoptosis~~ apoptosis in the Hela cells by chloroform fraction of seed ~~Extracts~~ extracts of *Nigella sativa,. Cancer Cell International.*9:29.

Shangari, N., O'Brien, P.J. (2006). Catalase ~~Activity~~ activity ~~Assays~~assays. Current ~~Protocols~~ protocols in ~~Toxicology~~toxicology.

Skrzydlewska, E., Stankiewiez, A., Sulkowska, M., Sulkowski, S. and Kasacka, I. (2001). Antioxidant ~~Status~~ status and ~~Lipid~~ lipid ~~Peroxidation~~ peroxidation in ~~Colorectal~~ colorectal ~~Cancer~~cancer. *J. Toxicol .Environ. Health*.64:213-22.

~~Skrzydlewska, E., Stankiewiez, A., Sulkowska, M., Sulkowski, S. and Kasacka, I. (2001). Antioxidant Status and Lipid Peroxidation in Colorectal Cancer.*J. Toxicol .Environ. Health*.64:213-22.~~

Sulistyoningrum, E., Rachmani, E.P.N., Baroroh,H.N., Rujito, L. (2017) *Annona muricata* ~~Leaves~~ leaves ~~Extract~~ extract ~~Reduce~~ reduce ~~Proliferative~~ proliferative ~~Indexes~~ indexes ~~And~~ and ~~Improve~~ improve ~~Histological~~ histological ~~Changes~~ changes ~~In~~ in ~~Rat's~~ rat's ~~Breast~~ breast ~~Cancer~~cancer. *J App PharmSci,* ; 7 (01): 149-155

Tietz,N.W. (1976).Fundamentals of ~~Clinical~~ clinical ~~Chemistry~~chemistry, W.B.,Saunders Co., Philadelphia,

Vander Heiden, M.G. (2011). Targeting ~~Cancer~~ cancer ~~Metabolism~~metabolism. A ~~Therapeutic~~ therapeutic window opens. *National Rev. Drug. Discov.* 10:671-684.

Formatted: Font: Italic

UNDER PEER REVIEW

Veech, R. L., Chance, B., Kashiwaya, Y., Lardy, H.A. and Cahill, G. F. (2001). Ketone bodies, potential therapeutic uses. *IUBMB life*.51(4): 241-247

Venkateshwarlu E., Swathi G., Sharvana B. B. S., Shiva K. R., Venkateshwar R. J. and Umasankar K. (2014). Evaluation of ~~Anticancer~~ anticancer ~~Activity~~ activity of *Annona muricata* in 1, 2-~~Dimethyl~~ dimethyl ~~Hydrazine~~ hydrazine ~~Induced~~ induced ~~Colon~~ colon ~~Cancer~~cancer. *World Appl. Sci. J.* 32; 444-450 10.5829/idosi.wasj.2014.32.03.85109.

Vijayameena, C., Subhashini, G., Loganayagi, M. and Ramesh, B. (2013). Phytochemical screening and ~~Assessment~~ assessment of ~~Antibacterial~~ antibacterial ~~Activity~~ activity for the ~~Bioactive~~ bioactive compounds in *Annona muricata. Int. J. Curr. Microbial.App.Sci.* 2(1): 1-8.

Warburg, O. (1956). "On the ~~Origin~~ origin of ~~Cancer~~ cancer ~~Cells~~cells".*Science*.123 (3191): 309-314.

Wilder, R. M. (1921). The ~~Effect~~ effect on ~~Ketonemia~~ ketonemia on the course of epilepsy. *Mayo.Clin. Bull.* 2:307

Williamson, D.H., Mellanby, J. and Krebs, H.A.(1962). Enzymatic ~~Determination~~ determination of D (-) –b-~~Hydroxybutyric~~ hydroxybutyric acid and ~~Acetoacetic~~ acetoacetic ~~Acid~~ acid in ~~Blood~~blood. *Biochem. J*.82:90-96.

Yang, C. R., Ou, Y. C. and Kuo, J. H. (1997).Intracellular ~~Glutathione~~ glutathione ~~Content~~ content of ~~Urothelial~~ urothelial ~~Cancer~~ cancer in ~~Correlation~~ correlation to ~~Chemotherapy~~ chemotherapy ~~Response~~response. *Cancer. Lett*.119:157-162.

Formatted: Font: Italic

UNDER PEER REVIEW

Zeng, L., Wu, F. E., Oberlies, N. H., McLaughun, J. L., Sastrodihadjo. S. (1996). Five ~~New~~ new ~~Monotetrahydrofuran~~ monotetrahydrofuran ~~Ring~~ ring ~~Acetogenins~~ acetogenins ~~From~~ from the leaves of *Annona muricata*. *J .Nat. Prod.* 59(11): 1035-1042.

UNDER PEER REVIEW