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Leaf Chlorophylls and Carotenoids Status and their correlation with storage root weight of Some Local and Exotic Sweetpotato Genotypes

ABSTRACT

The investigation was carried out to characterize the chlorophyll components and carotenoids of leaves of some local and exotic genotypes of sweetpotato namely Local-1, Local-2, Local-5, Local-8, Exotic-1, Exotic-2, Exotic-4 and BARI SP-4 and their effect on production of total dry matter and storage roots dry weight during November 2016 to April 2017 at farmer's field of Dashpara village of Sylhet Sadar Upazila, Sylhet, Bangladesh. The experiment was laid out in Randomized Complete Block Design with three replications. Fresh leaves of 5-6th position from the top of vine were collected from the research field into polybag with proper tagging and brought to the laboratory in the morning of 30, 60, 90 and 120 days after planting (DAP). Collected leaves were washed, wiped out of excess water, cut into small pieces, mixed thoroughly, and 250 mg of leaf materials were taken in a mortar. Leaf materials were grinded finely by a pestle with 25 ml of cold 80% acetone for two minutes. Sample tubes were centrifuged for 10 minutes. The homogenate was filtered and made up to 25 ml with cold 80% acetone. The centrifuged samples were incubated in dark for half an hour. The optical density (OD) for each solution was measured at 663, 645 and 440.5 nm against 80% acetone as blank in one cm cell of spectrophotometer. Triplicate estimation was done for each sample. Chemical analyses were performed at Regional Laboratory of Soil Resource Development Institute, Sylhet. Statistical analyses was done using MSTATC software following analysis of variance technique and Duncan's Multiple Range Test. Results showed that chlorophyll-a gradually increased up to 60 DAP in all genotypes, thereafter it continued only in Exotic-4, Exotic-3 and Local-1 up to 90 DAP. The highest amount of chlorophyll-a (10.27±0.45 mg 100 gfw⁻¹) was in Local-1 at 90 DAP. The highest amount of chlorophyll-b was in Exotic-3 (19.13±0.53 mg 100 gfw⁻¹) followed by Local-1 (16.85±0.50 mg 100 gfw⁻¹) at 30 DAP. Carotenoids content in leaves of all genotypes increased gradually up to 90 DAP and thereafter decreased except Exotic-4. The highest carotenoids was in Exotic-3 (10.78 mg 100 gfw⁻¹) followed by Local-1 (10.13 mg 100 gfw⁻¹) at 90 DAP. At 120 DAP, the highest storage roots weight was in Local-8 (232.40±5.97 g plant⁻¹), followed by Local-1 (187.50±5.23 g plant⁻¹). Chlorophylls and carotenoids had no significant effect on total dry matter and storage roots dry weights at 30 DAP. At 120 DAP, all chlorophyll components and carotenoids had positive correlation with total dry matter (TDM) and storage roots dry weights. Genotypes Local-1, Local-8 had the higher chlorophylls while Exotic-3, Local-1 and Local-8 had the higher carotenoids. Genotypes Local-1 and Local-8 showed the highest storage roots dry weight.

Keywords: Chlorophylls, carotenoids, exotic genotype, local genotype, yield, correlation

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1. INTRODUCTION

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Sweetpotato (Ipomoea batatas L. Lam.) is a dicotyledonous plant of the family Convolvulaceae. It is perennial in nature but it is grown as an annual crop. The plants bear adventitious roots which enlarge near the stem and form edible storage roots. In Bangladesh it is produced at about 0.761 million ton on about 0.045 million ha of land with an average yield of 16.91 t ha⁻¹ [1]. It is characterized by low production, yield and storage root quality compared to Japan, Senegal and Israel [2]. However, it is easy to grow and capable of growing under adverse weather and soil conditions. It requires low input and less management practices [3]. It is a very efficient food crop and produces more dry matter, protein and minerals per unit area in comparison to cereals [4]. Storage roots are rich source of energy, several minerals and micronutrients [5] and leaves are rich in vitamin B, beta-carotene, iron, calcium, zinc and protein [6].

The yield of sweetpotato depends on the production of assimilates (source) and its accumulations (sink). Storage roots (number and weight) are predominant sink whereas leaves and tender vines are the source. The photosynthetic rate and the leaf area are regarded as the source potential. Leaves take part in the production of assimilates. The leaves of plants contain chlorophylls (Chlorophyll-a, Chlorophyll-b) and carotenoids. Chlorophyll-a possesses a green-blue color while chlorophyll-b possesses green-yellow color [7]. Chlorophyll with the pigments has a central role in light harvesting, photosystem protection, and other growth functions [8, 9]. Carotenoids participate in harvesting light energy for photosynthesis [10]. They are also involved in the defense mechanism against oxidative stress [11], and play an essential role in the dissipation of excess light energy and provide protection to reaction centers [12, 13].

Recent studies showed that chlorophyll and carotenoids have positive effect on human health. Chlorophyll is often referred to as the green blood of plants due to the identical molecular structure with hemoglobin with only difference in center atom (iron or magnesium). This similarity makes chlorophyll so important to our health, it improve digestive, immune and detoxification systems of human body [14]. Leaves contain phenols, flavonoids, β -carotene, anthocyanin, and caffeoylquinic acid derivatives [15]. Carotenoids extract from leaves functions as a cheap natural yellow dye. It can be beneficial to human health compare to the artificial colouring dye [16].

There are many local sweetpotato genotypes available in Sylhet region and many of them are growing at the farmer's level sporadically. Rajput *et al.* [17] reported that the functional leaves may directly reflect to yield. Besides, for adaptive trial of improved sweetpotato cultivars developed by different countries and organizations, it is necessary to determine chlorophyll and carotenoids. Moreover, it is necessary to determine how these genotypes/cultivars can be made available to the farmers of Bangladesh. Therefore, an experiment was undertaken to characterize the chlorophyll components and carotenoides of leaves of some local and exotic genotypes of sweetpotato and their effect on production of total dry matter and storage roots dry weight.

2. METHODOLOGY

The experiment was carried out at farmer's field of Dashpara village of Sylhet Sadar Upazila, Sylhet during November 2016 to March 2017. It lies between 24°54′32.8″ to 24°54′33.5″ N and latitude and 91°56′ 59.5″ to 91°57′00.9″ E longitude. Texture of top soils was sandy loam (sand 55%, silt 42%, clay 3%). Well drained soil. Top soil (0-15 cm) is Top to sub soil is light brown to brown mottled grey colour. The soil is characterized as Bijipur soil series of Northern an Eastern Piedmont Plains (AEZ 22) in Bangladesh [18].

2.1 Planting Materials

67 Nine sweeetpotato genotypes/cultivars namely Local-1, Local-2, Local-5, Local-8, Exotic-1,

68 Exotic-2, Exotic-4 and BARI SP-4 were used as planting materials while BARI SP-4 was

69 check variety.

2.2 Experimental Procedure and Design

The experiment was set in a Randomized Complete Block Design (RCBD) with three replications. The experimental field was fertilized with manures and fertilizers as per soil test value: Cowdung =5000 kg, Urea =214 kg, TSP =171 kg, MoP =188 kg, Gypsum =56 kg, Zinc sulfate (Hepta) =10 kg, Solubor =3 kg, Magnesium sulfate =82 kg and dolomite =988 kg. Before final land preparation, half of urea and MoP, full of other fertilizers and cow dung were applied. Rest of Urea and MoP were applied as side dressing after 35 days of planting at earthing up operation. Soil reaction (pH) was corrected by dolomite application prior to 15 days of planting followed by bed preparation. Raised beds were prepared and cuttings of sweetpotato vines were planted in lines maintaining row to row 60 cm and plant to plant 30 cm. The unit plot size was of 4.8 m x 4.2 m with a block to block distance 1.0 m and plot to plot distance 0.6 m. Weeding was done as and when necessary. Irrigation was done at 30 and 60 days after planting (DAP).

2.3 Data Collection

Leaves were collected at 30, 60, 90 and 120 DAP and chlorophyll a, chlorophyll b, chlorophyll a/b ratio, total chlorophyll and carotenoids contents were estimated.

2.4 Sample Preparation and Chemical Analysis

Collected fresh leaves were cut into small pieces leaving away mid ribs, mixed thoroughly and 250 mg of leaf materials were taken in a mortar and small amount of sodium carbonate (Na_2CO_4) was added into it to check the degradation of pigments. Leaf materials were grinded finely by a pestle with 25 ml of cold 80% acetone for two minutes. Sample tubes were centrifuged for 10 minutes (Model- SORVALL Legend Micro 21R Centrifuge, Thermo Fisher Scientific, Germany). The homogenate was filtered through Whatman number 1 filter paper and made up to 25 ml with cold 80% acetone. The centrifuged samples were incubated in dark for half an hour. The optical density (OD) for each solution was measured at 663, 645 and 440.5 nm (Model T80+, UV/VIS Spectrometer, PG Instruments Ltd., UK) against 80% acetone as blank in one cm cell. The amount of chlorophyll-a and chlorophyll-b were determined by using specific absorption coefficient of McKinney [19] and the formula of Maclachalan and Zalik [20], Duxbury and Yentsch [21]. The amount of carotenoids was determined by the equation of Holm [22].

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i) C_a = \frac{(12.3 \times D663 - 0.86 \times D645)V}{d \times 1000 \times W} mg/g fresh leaf [20]

ii) C_b = \frac{(19.3 \times D645 - 3.6 \times D663)V}{d \times 1000 \times W} mg/g fresh leaf [21]
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             iii) C_c = 4.695 \times D440.5 - 0.268 C (a+b) [22]
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             iv) Total chlorophyll (mg g fresh leaf<sup>-1</sup>) = Chlorophyll a + Chlorophyll b
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            v) Ratio of chlorophyll-a and Chlorophyll-b = \frac{\text{Chlorophyll a}}{\text{Chlorophyll b}}
109
110
            Ca = Chlorophyll a (mg g fresh leaf<sup>-1</sup>)
111
112
            Cb = Chlorophyll b (mg g fresh leaf<sup>-1</sup>)
            D = Optical density (O.D.) at wave length indicated
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114 V = Final volume (ml)

115 W = Fresh weight of leaf materials used (g)

116 d = Length of light path (cm)

117 Cc = Concentration of carotenoids in µg ml⁻¹, which was then converted into mg
118 q⁻¹ fresh leaf

2.5 Statistical Analysis of Data

The data were analyzed using Analysis of Variance (ANOVA) technique through MSTATC package. Comparative analysis of the results was done using Duncan's Multiple Range Test (DMRT) at 1% level of significance. A p-value p \leq 0.01) was considered statistically significant.

3. RESULTS AND DISCUSSION

3.1 Chlorophyll Content in Leaves

Chlorophyll-a content (mg 100 gfw⁻¹) in leaves of all genotypes showed that it gradually increased up to 60 DAP, after that it increased sharply in Exotic-4, Exotic-3 and Local-1, and decreased in Local-2, Local-5 and Exotic-1 up to 90 DAP (Fig. 1). After 90 days of planting, chlorophyll-a content decreased sharply up to 120 DAP. Local-1 produced the highest amount of chlorophyll-a (5.63±0.08) at 30 DAP, increased to 8.80±0.58 at 60 DAP, 10.27±0.45 at 90 DAP and finally reduced to 3.45±0.09 at 120 DAP.

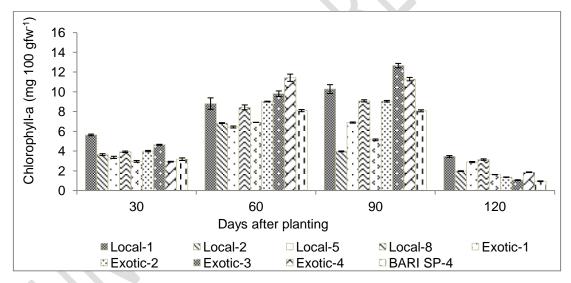


Fig. 1. Effect of genotype on the chlorophyll-a content in leaves of sweetpotato at different days after planting (DAP)

** $P \le 0.01$, Mean \pm SEM=Mean values \pm Standard error of means,n=3, gfw⁻¹=g fresh weight

Chlorophyll-b content in leaves (mg 100 gfw⁻¹) had significant variations (Fig. 2). At initial stage, the amount of chlorophyll-b was higher in all of the genotypes and thereafter decreased dramatically. At 30 DAP, the highest amount of chlorophyll-b (mg 100 gfw⁻¹) was found in Exotic-3 (19.13±0.53) followed by Local-1 (16.85±0.50). At 60 DAP, the highest amount of chlorophyll-b was in Local-1 (5.57±0.07) followed by Local-2 (5.37±0.22) and Local-8 (4.76±0.30) and the lowest was in Exotic-1 (1.94±0.04). At 90 DAP, the highest amount was in Local-1 (5.60±0.01) followed by Exotic-3 (4.54±0.17) and the lowest was in Exotic-1 (1.62±0.03). At 120 DAP, the highest amount was in Local-8 (4.55±0.06) followed by Local-1 (4.26±0.15) and the lowest was in Exotic-3 (1.41±0.10). The ratio of chlorophyll-a to chlorophyll-b in higher plants is approximately 3:1 [7].

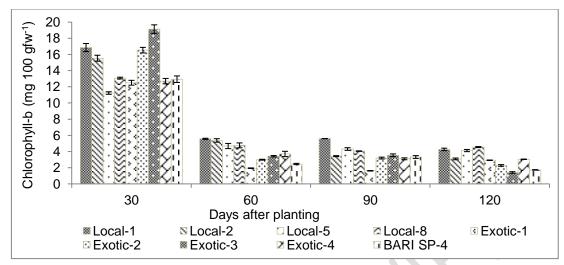


Fig. 2. Effect of genotype on the chlorophyll-b content in leaves of sweetpotato at different DAP

** $P \le 0.01$, Mean \pm S.E.M = Mean values \pm Standard error of means, n=3.

As like chlorophyll-a and chlorophyll-b, the ratio of them was also very low at initial stage and as days passes the ratio was increased up to 60 DAP (Fig. 3). From 60-90 DAP, ratio of Local-1, Local-5, Local-8 and Exotic-4 increased whereas ratio of Local-2, Exotic-1, Exotic-2, Exotic-3 and BARI SP-4 decreased gradually. After 90 days of planting, ratio decreased harshly. The highest ratio (3.65±0.17) was in Exotic-4 and the lowest was in Local-2 (1.15±0.03) at 90 DAP. The chlorophyll a/b ratio of Local-1 was seen to be 0.34±0.01 at 30 DAP, 1.58±0.11 at 60 DAP, 1.81±0.08 at 90 DAP and 0.81±0.04 at 120 DAP.

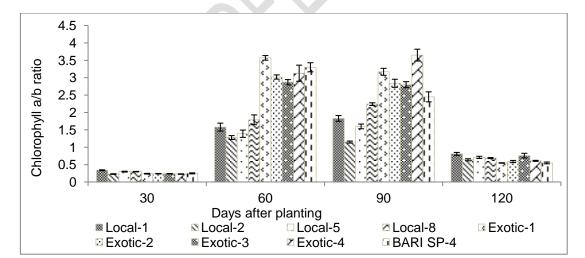


Fig. 3. Effect of genotype on the chlorophyll a/b ratio in leaves of sweetpotato at different DAP

** $P \le 0.01$, Mean \pm S.E.M = Mean values \pm Standard error of means, n=3.

Total chlorophyll consists of chlorophyll-a and chlorophyll-b varied significantly (Table 1). Initially although the total chlorophyll content (mg 100 gfw⁻¹) was high but it reduced with plant ages in all of the genotypes. After 30 days of planting, the highest total chlorophyll content was in Exotic-3 (23.76±0.56) followed by Local-1 (22.48±0.46) and the lowest in

Local-5 (14.58±0.32). After 60 days of planting, the highest total chlorophyll content was in Exotic-4 (15.12±0.60) followed by Local-1 then Exotic-3 and Local-8. The lowest was in Exotic-1 (8.84±0.04). After 90 days of planting, the highest total chlorophyll content was in Exotic-3 (17.18±0.34) followed by Local-1 (15.87±0.44) and the lowest were in Exotic-1 (6.75±0.08). At final harvest (120 DAP), Local-1 (7.71±0.13 mg) and Local-8 (7.68 mg) were showed similar and the highest content of total chlorophyll.

Table 1. Total chlorophyll content in leaves of sweetpotato genotypes at different DAP

Genotypes	Total chlorophyll content in leaves at different DAP (mg 100 gfw ⁻¹)					
	30	60	90	120		
Local-1	22.48±0.46 b	14.37±0.55 ab	15.87±0.44 b	7.71±0.13 a		
Local-2	19.16±0.33 d	12.20±0.20 cd	7.38±0.06 g	5.04±0.09 c		
Local-5	14.58±0.14 g	11.11±0.28 de	11.19±0.15 f	7.00±0.12 b		
Local-8	16.99±0.09 e	13.18±0.28 bc	13.13±0.10 d	7.68±0.12 a		
Exotic-1	15.45±0.27 f	8.84±0.04 f	6.75±0.08 g	4.53±0.02 d		
Exotic-2	20.52±0.32 c	12.00±0.05 cd	12.22±0.10 e	3.63±0.09 e		
Exotic-3	23.76±0.56 a	13.22±0.3 bc	17.18±0.34 a	2.46±0.09 f		
Exotic-4	15.62±0.32 f	15.12±0.60 a	14.38±0.15 c	4.89±0.03 c		
BARI SP-4	16.10±0.36 f	10.54±0.10 e	11.39±0.15 ef	2.67±0.03 f		
CV (%)	1.51	4.23	3.10	2.45		
LSD _{.01}	0.657	1.239	0.902	0.292		

Figures (Mean \pm S.E.M) in a column having different letters differ significantly at 1% level of significance by DMRT

The variations in chlorophyll-a, chlorophyll-b, and their corresponding a/b ratio and total chlorophyll were probably due to genotypic, fertilization as well as growth stages of leaves. The results corroborate with findings of Katayama and Shida [23] and Yooyongwech *et al.* [24]. Katayama and Shida [23] reported chl a, chl b and ratio a/b were 69.1 mg 100 gfw⁻¹, 23.5 mg 100 gfw⁻¹ and 2.949, respectively in the leaves of 6th position in the sweetpotato vine. They reported that the contents of chlorophyll a and b will change in their absolute amount and also in their ratio a/b according to the kind of materials or to the different developmental stages as well as fertilizers, chemicals, moisture and other environments. They added that the change of chlorophyll contents was observed corresponding to the developmental stages of leaves in sweetpotato. Rashid [25] established that the content of chlorophyll-a, chlorophyll-b, and their ratio were influenced by the cultivar.

On the other hand, Yooyongwech *et al.* [24] reported the chlorophyll a, chlorophyll b, total chlorophyll contents and ratio Chl a: Chl b in three genotypes of sweetpotato grown under well watering in the pot culture were ranged 24.07-33.46 mg 100 gfw⁻¹, 11.75-14.43 mg 100 gfw⁻¹, 36.95-47.89 mg 100 gfw⁻¹ and 1.71-2.32 mg 100 gfw⁻¹, respectively. They reported that the variation of the chlorophyll contents of the present result was perhaps due to genetic makeup.

3.2 Carotenoids Content in Leaves

Carotenoids content (mg 100 gfw⁻¹) of all genotypes increased gradually up to 90 DAP and thereafter decreased except Exotic-4 (Fig. 4). At 90 DAP, the highest carotenoids was in Exotic-3 (10.78 mg) followed by Local-1 (10.13 mg) and the lowest was in Exotic-1 (7.93 mg). Overall, all of the genotypes were found better over the check variety BARI SP-4 in carotenoids production except Exotic-1.

The variations in carotenoids among the genotypes were due to the influence of genotypic and/or environmental conditions and developmental stages of leaves. Woolfe [4] reported carotenoids in sweetpotato leaves ranged from 0.38 to 7.24 mg 100 gfw⁻¹ while Motsa *et al.*

[26] reported that carotenoids and total chlorophyll content of edible leaves were ranged from 90 to 390 mg 100 gdw⁻¹ and 1.54 to 4.47 mg 100 gdw⁻¹, respectively. They added that total carotenoids and chlorophyll content in leaves were significantly affected by environmental conditions.

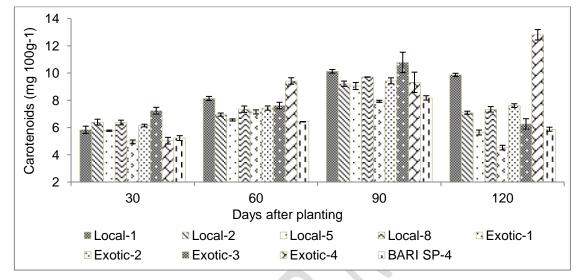


Fig. 4. Effect of genotype on the carotenoids content in leaves of sweetpotato at different DAP

** $P \le 0.01$, Mean \pm S.E.M = Mean values \pm Standard error of means, n=3.

3.3 Total Dry Matter

Total dry matter (TDM) increased gradually up to 60 DAP and thereafter increased very rapidly upto 120 DAP in all of the genotypes (Table 2). At 120 DAP, the highest TDM was in Local-8 (327.10±5.52 g plant⁻¹) followed by Local-1 (292.30±5.65 g plant⁻¹), and the lowest was in Exotic-3 (116.90±1.36 g plant⁻¹).

Table 2. Effect of genotypes on the total dry matter of sweetpotato at different DAP

Genotypes		Total dry matter (g plant ⁻¹)					
	30	60	90	120			
Local-1	23.03±1.22 a	34.87±1.16 a	100.30±1.22 b	292.30±5.65 b			
Local-2	17.97±0.50 b	32.38±0.43 b	93.82±0.96 c	232.90±2.39 c			
Local-5	12.08±0.46 c	23.42±0.52 d	74.07±1.64 e	170.90±2.87 e			
Local-8	15.32±0.28 b	32.06±0.31 b	118.70±2.97 a	327.10±5.52 a			
Exotic-1	15.28±0.49 b	22.72±0.43 d	56.85±0.71 fg	134.20±1.57 g			
Exotic-2	16.27±0.87 b	28.19±0.70 c	77.56±1.83 e	157.10±0.56 f			
Exotic-3	9.11±0.22 d	16.98±0.50 e	51.68±1.66 g	116.90±1.36 h			
Exotic-4	16.53±0.52 b	24.66±0.22 d	62.76±1.15 f	175.80±1.50 e			
BARI SP-4	11.80±0.09 c	23.99±0.33 d	85.46±1.31 d	209.10±3.13 d			
CV%	7.20	3.89	3.32	2.69			
Lsd .01	2.622	2.468	6.350	12.970			

Figures (Mean ± SEM) in a column having different letters differ significantly at 1% level of significance by DMRT

Hossain and Islam [27] reported that total dry weights of 10 sweetpotato genotypes increased up to 165 DAP. Nandi and Sen [28] reported that the total biomass yield increased linearly upto 120 DAP except two genotypes. Nair and Nair [29] reported a linear increase in TDM. Mannan *et al.* [30] established that TDM increased rapidly from 90 to 150 DAP. Haque [31] reported that TDM had a linear growth phase that continued until about 120 DAP. Watson [32] stated that TDM production of a crop is dependent on the source and its activities as well as the length of its growth period, during which photosynthesis continues.

3.4 Storage Root Dry Weight

Storage roots dry weight increased gradually up to 90 DAP and then increased sharply up to 120 DAP (Table 3). After 30 days of planting, storage roots appeared only in Local-8 (0.65±0.09 g plant⁻¹) and check variety BARI SP-4 (1.88±0.08 g plant⁻¹). After 60 days of planting, all of the genotypes initiated storage roots, except Exotic-1 and Exotic-4. After 90 days of planting, all of the genotypes initiated storage roots. After 120 days of planting, the highest weight was in Local-8 (232.40±5.97 g plant⁻¹), followed by Local-1 (187.50±5.23 g plant⁻¹) and the lowest weight was in Exotic-1 (54.05±1.11 g plant⁻¹).

Table 3. Effect of genotypes on the storage roots dry weight of sweetpotato at different DAP

Genotypes	Storage root dry weight (g plant ⁻¹)					
	30	60	90	120		
Local-1	0.00±0.00 c	2.73±0.04 de	31.13±0.46 c	187.50±5.23 b		
Local-2	0.00±0.00 c	4.30±0.14 c	29.88±0.80 cd	152.90±1.61 c		
Local-5	0.00±0.00 c	2.90±0.05 d	25.08±1.36 de	98.28±1.53 d		
Local-8	0.65±0.09 b	7.97±0.02 b	60.09±2.47 a	232.40±5.97 a		
Exotic-1	0.00±0.00 c	0.00±0.00 g	13.07±0.91 f	54.05±1.11 f		
Exotic-2	0.00±0.00 c	2.57±0.09 e	29.18±0.91 cd	86.42±1.31 d		
Exotic-3	0.00±0.00 c	1.66±0.06 f	22.80±1.62 e	68.98±0.32 e		
Exotic-4	0.00±0.00 c	0.00±0.00 g	8.720±0.15 f	93.39±1.55 d		
BARI SP-4	1.88±0.08 a	8.27±0.08 a	53.24±0.22 b	144.20±3.14 c		
CV%	17.48	3.48	7.37	4.12		
Lsd .01	0.1067	0.2822	5.337	12.22		

Figures (Mean ± SEM) in a column having different letters differ significantly at 1% level of significance by DMRT

The above results corroborate with the findings of Oswald *et al.* [33] where the storage root dry matter of sweetpotato increment followed a sigmoid pattern, and Nair and Nair [29] while they reported a linear increase in storage root dry matter and the increase in storage root dry matter was the maximum during 48 to 161 days of planting.

3.5 Correlation of Chlorophyll and Carotenoids with Yield of Sweetpotato

At 30 DAP chlorophylls and carotenoids had no significant effect on total dry matter and storage roots dry weight (Table 4). However, Chlorophyll-a was highly correlated with chlorophyll-b, total chlorophyll, chlorophyll a /b ratio and carotenoids whereas chlorophyll-b correlated with total chlorophyll and carotenoids.

After 60 days of planting chlorophyll-b had positive significant correlation with TDM ($r = 0.645^{**}$) while chlorophyll a/b ratio correlated significantly but negatively with TDM ($r = 0.587^{**}$) (Table 5). Carotenoids had significantly negative correlation with storage roots dry weight, however had positive correlation with chlorophyll-a and total chlorophylls. Similar

correlation effect was observed at 90 DAP while carotenoids had insignificantly positive correlation with storage root dry weight (Table 6).

After 120 days of planting, all chlorophyll components and carotenoids had positive correlation with TDM and storage roots dry weight (Table 7) while chlorophyll-a, chlorophyll-b and total chlorophylls had significant correlation with them. It was also indicated that Chorophyll-a had no effect on storage roots dry weight upto 90 DAP while chlorophyll-b and total chlorophyll upto 60 DAP. Chlorophyll-b had positive correlation with TDM from the beginning while chlorophyll-a, total chlorophyll and chlorophyll a/b ratio was negative or insignificant upto 120 DAP.

Table 4. Correlation of chlorophyll and carotenoids with total dry matter and storage root dry weight (g plant⁻¹) at 30 DAP

	TDM	SDW	Chl-a	Chl-b	Total chl Chl a/b ratio
SDW	-0.307				
Chl-a	0.375	-0.250			
Chl-b	0.099	-0.282	0.722**		
Total chl	0.178	-0.290	0.836**	0.983**	
Chl a/b ratio	0.333	-0.049	0.612**	-0.096	0.086
Carotenoids	-0.187	-0.218	0.514**	0.731**	0.715** -0.011

TDM = Total dry matter (g plant⁻¹), SDW = Storage root dry weight (g plant⁻¹), Chl-a = Chlorophyll-a, Chl-b = Chlorophyll-b, Total chl = Total chlorophyll, Chl a/b ratio = Ratio chlorophyll-a and chlorophyll-b

Table 5. Correlation of chlorophyll and carotenoids with total dry matter and storage root weight (g plant⁻¹) at 60 DAP

	TDM	SDW	Chl-a	Chl-b	Total chl	Chl a/b ratio
SDW	0.349					
Chl-a	-0.152	-0.260				
Chl-b	0.645**	0.179	-0.093			
Total chl	0.301	-0.094	0.754**	0.583**		
Chl a/b ratio	-0.587 ^{**}	-0.275	0.396*	-0.936 ^{**}	-0.295	
Carotenoids	0.089	-0.522	0.775**	0.138	0.723**	0.165

TDM = Total dry matter (g plant⁻¹), SDW = Storage root dry weight (g plant⁻¹), Chl-a = Chlorophyll-a, Chl-b = Chlorophyll-b, Total chl = Total chlorophyll, Chl a/b ratio = Ratio chlorophyll-a and chlorophyll-b

Table 6. Correlation of chlorophyll and carotenoids with total dry matter and storage root weight (g plant⁻¹) at 90 DAP

	TDM	SDW	Chl-a	Chl-b	Total chl	Chl a/b ratio
SDW	0.783					
Chl-a	-0.171	-0.037				
Chl-b	0.397	0.282	0.487**			
Total chl	-0.010	0.061	0.960**	0.712**		
Chl a/b ratio	-0.571 ^{**}	-0.391 [*]	0.478 [*]	-0.510 ^{**}	0.221	
Carotenoids	0.088	0.004	0.610**	0.713	0.719	-0.165

TDM = Total dry matter (g plant⁻¹), SDW = Storage root dry weight (g plant⁻¹), Chl-a = Chlorophyll-a, Chl-b = Chlorophyll-b, Total chl = Total chlorophyll, Chl a/b ratio = Ratio chlorophyll-a and chlorophyll-b

Table 7. Correlation of chlorophyll and carotenoids with total dry matter and storage root weight (g plant⁻¹) at 120 DAP

	TDM	SDW	Chl-a	Chl-b	Total chl	Chl a/b ratio
SDW	0.984**					
Chl-a	0.715**	0.634**				
Chl-b	0.686**	0.591**	0.950**			
Total chl	0.708**	0.618**	0.985**	0.990**		
Chl a/b ratio	0.303	0.306	0.563**	0.301	0.425*	
Carotenoids	0.268	0.202	0.252	0.230	0.243	0.148

TDM = Total dry matter (g plant⁻¹), SDW = Storage root dry weight (g plant⁻¹), Chl-a = Chlorophyll-a, Chl-b = Chlorophyll-b, Total chl = Total chlorophyll, Chl a/b ratio = Ratio chlorophyll-a and chlorophyll-b

3.4 Contribution to the Existing Knowledge/Implication of the Study

From the study it was observed that chorophyll-a had no effect on storage roots dry weight production upto 90 days of planting while chlorophyll-b and total chlorophyll had no effect before 60 days of planting. After 120 days of planting storage root dry weight production was influenced significantly by chlorophyll-a, chlorophyll-b and total chlorophyll. On the other hands, carotenoids had negative effect on storage roots dry weight at 60 DAP and started to positive effect at 90 DAP which indicates that carotenoids accumulation into leaves that would finally translocate to the storage roots, started after 90 days of planting. Based on the information we can easily select the suitable sweetpotato genotypes and perform the crop management practices.

4. CONCLUSION

Chlorophyll-a content in leaves of sweetpotato genotypes decreased after 60 days of planting while chlorophyll-b decreased after 30 days of planting. Carotenoids content in leaves were decreased after 90 days of planting. At 120 DAP, genotypes Local-1, Local-8 had the higher chlorophylls while Exotic-3, Local-1 and Local-8 had the higher carotenoids. Genotypes Local-8, Local-1 and Local-2 had the higher storage roots dry weight at 120 DAP. Chorophyll-a had no effect on storage roots dry weight upto 90 DAP where chlorophyll-b and total chlorophyll had come to effect after 60 DAP. At 120 DAP, storage root dry weight was influenced significantly by chlorophyll-a, chlorophyll-b and total chlorophyll. Genotypes Local-1 and Local-8 showed the highest dry matter yield performance.

ACKNOWLEDGEMENTS

This work was supported in part by a grant from Bangladesh University Grants Commission, Dhaka.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- [1] Ministry of Agriculture, Govt. of Bangladesh. Crop situation in Bangladesh. 2017. Available: www.moa.gov.bd, <a href="https://https:/
- [2] Food and Agriculture Organization. FAO Statistical Year Book of the United Nations, Regional Office for Asia and the Pacific Bangkok. 2014; 176. Available at: www.fao.org

- [3] Kozai T, Kubota C and Kitaya Y. Sweetpotato technology for solving the global issues on food, energy, natural resources and environment in the 21st century. Environ. Control in Biol. 2006: 34:105–114.
- 338 [4] Woolfe JA. Sweetpotato: An untapped food resource, Cambridge University Press, Cambridge, UK. 1992; pp. 643.

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355

359

362 363

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365

366 367

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369

370 371

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- [5] Laurie SM, Faber M, van Jaarsveld PJ, Laurie RN, du Plooy CP, Modisane PC. Beta-carotene yield and productivity of orange-fleshed sweetpotato (*Ipomoea batatas* (L.) Lam.)
 as influenced by irrigation and fertilizer application treatments. Sci. Hortic. 2012; 142:180-184.
- [6] Islam S. Nutritional and medicinal qualities of sweetpotato tops and leaves. Cooperative
 Extension Program, University of Arkansas, Pine Bluff, Chicago, USA. 2014.
- [7] Arnon DI. Copper enzyme in isolated chloroplast polyphenol oxidase in *Beta vulgaris* (L.). Plant Physiology. 1949; 24:1-5.
- 352 [8] Abramavicius D, Valkunas L. Role of coherent vibrations in energy transfer and conversion in photosynthetic pigment-protein complexes. Photosynth. Res. 2016; 127, 33–47.
- [9] Batjuka A, Skute N, Petjukevics A. The influence of anthocyanin on pigment composition and functional activity of photosynthetic apparatus of *Triticum aestivum* L. under high temperature. Photosynthetica. 2016; 55: 1–14.
- [10] Zakar T, Laczko-Dobos H, Toth TN, Gombos Z. Carotenoids assist in cyanobacterial photosystem II assembly and function. Front. Plant Sci. 2016; 7:295.
 - [11] Campos MD, Nogales A, Cardoso HG, Campos C, Grzebelus D, Velada I, Arnholdt-Schmitt B. Carrot plastid terminal oxidase gene (dcptox) responds early to chilling and harbors intronic pre-mirnas related to plant disease defense. Plant Gene. 2016; 7:21–25.
 - [12] Santabarbara S, Casazza AP, Ali K, Economou CK, Wannathong T, Zito F, Redding KE, Rappaport F, Purton S.The requirement for carotenoids in the assembly and function of the photosynthetic complexes in chlamydomonas reinhardtii. Plant Physiol. 2013; 161:535–546.
 - [13] Nagy L, Kiss V, Brumfeld V, Osvay K, Borzsonyi A, Magyar M, Szabo T, Dorogi M, Malkin. Thermal effects and structural changes of photosynthetic reaction centers characterized by wide frequency band hydrophone: Effects of carotenoids and terbutryn. Photochem. Photobiol. 2015; 91:1368–1375.
- [14] Kopsell DA, Kopsell DE, Curran-Celentano J. Carotenoid and chlorophyll pigments in sweet basil grown in the field and greenhouse. HortScience. 2005; 40 (5):1230–1233.
 377
- [15] Rumbaoa RG, Cornago DFand Geronimo IM Phenolic content and antioxidant capacity of Philippine sweetpotato(*Ipomoea batatas*) varieties. Food Chem. 2009; 113:1133–1138.
- 381 [16] Hue SM, Boyce AN, Chandran S. Influence of growth stage and variety on the pigment levels in *Ipomoea batatas* (sweetpotato) leaves. African Journal of Agricultural Research. 2011; 6(10):2379-2385.
- In [17] Rajput A, Rajput SS, Jha G. Physiological parameters leaf area index, crop growth rate, relative growth rate and net assimilation rate of different varieties of rice grown under

387 different planting geometries and depths in SRI. Int. J. Pure App. Biosci. 2017; 5(1):362-367. doi: http://dx.doi.org/10.18782/2320-7051.2472

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425

435

438

- 390 [18] SRDI (Soil Resource Development Institute). Land and soil statistical appraisal book of 391 Bangladesh, 1st edn., Ministry of Agriculture, Govt. of Bangladesh. Dhaka. 2010; pp.11-103. 392
- [19] Mackinney G. Criteria for purity of chlorophyll preparations. Journal of Biological Chemistry. 1940; 132:91-109.
- [20] Maclachlan S, Zalik S Plastid structure, chlorophyll concentration, and free amino acid
 composition of a chlorophyll mutant of barley. Canadian Journal of Botany. 1963; 41: 1053 1062.
- [21] Duxbury AC, Yentsch CS. Plant and pigment monography. Journal of Air Pollution and Control Assessment. 1956; 16:145-150.
- 403 [22] Holm G. Chlorophyll mutations in Barley. Acta Agric. (Scand.). 1954; 4:457-461.
- 405 [23] Katayama Y, Shida S. Studies on the change of chlorophyll a and b contents due to projected materials and some environmental conditions. Cytologia. 1970; 35:171-180.
- 408 [24] Yooyongwech S, Samphumphuang T, Theerawitaya C, Chaum S. Physio-morphological responses of sweetpotato [*Ipomoea batatas* (L.) Lam.] genotypes to water-deficit stress. J. Plant Omics. 2014; 7(5):361-368.
- 412 [25] Rashid MMHA. Comparative growth and yield studies of some exotic and local cultivars 413 of sweetpotato. M.Sc. (Agriculture) Thesis, Dept. of Crop Botany, Bangladesh Agricultural 414 University, Mymensingh. 2002. 415
- 416 [26] Motsa NM, Modi AT, Mabhaudhi T. Influence of agro-ecological production areas on 417 antioxidant activity, reducing sugar content, and selected phytonutrients of orange-fleshed 418 sweetpotatocultivars. Food Sci. Technology (Cam-606ea batatas pinas). 2015; 35(1):32-37. 419 [27] Hossain MAS, Islam AFMS. Comparative study on dry matter partitioning in five exotic 420 genotypes and five local varieties of sweetpotato. J. Sher-e-Bangla Agric. University. 2010; 421 4(1):24-30.
- [28] Nandi S, Sen H. Evaluation of sweetpotato (*Ipomoea batatas* L. Lam.) cultivars under late planted situation. Root Crops J. 1998; 24(1):73-77.
- 426 [29] Nair GM, Nair VM. Influence of irrigations and fertilizers on the growth attributes of sweetpotato, *Journal of Root Crops*. 1995: 21:17-23.
- [30] Mannan MA, Bhuiyan MKR, Quasem A, Rashid MM, Sidd. ique MA. Study on the growth and partitioning of dry matter in sweet potato. *J. Root Crops.* 1992; 18:1–5.
- 431 [31] Haque MM. Productivity of Maize/sweetpotato intercropping in relation to planting 432 system, population density and application of nitrogen and potassium fertilizers, PhD 433 Dissertation, Dept. of Agronomy, Institute of Post-Graduate Studies in Agriculture, Salna, 434 Gazipur-1703, Bangladesh.1995; pp.164.
- 436 [32] Watson DJ. The dependence of net assimilation rate on leaf area index. Ann. Bot. 1958; 437 22:37-54.

[33] Oswald A, Alkamper J, Midmore DJ. The effect of different shade levels on growth and tuber yield of sweetpotato, 1. Plant development. *Journal Agron. Crop Sci.* 1994; 175:99-107.

